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(54) Title: ISOLATED HUMAN PHOSPHATASE PROTEINS, NUCLEIC ACID MOLECULES ENCODING HUMAN PHOSPHATASE PROTEINS, AND USES THEREOF

(57) Abstract: The present invention provides amino acid sequences of peptides that are encoded by genes within the human genome, the phosphatase peptides of the present invention. The present invention specifically provides isolated peptide and nucleic acid molecules, methods of identifying orthologs and paralogs of the phosphatase peptides, and methods of identifying modulators of the phosphatase peptides.

ISOLATED HUMAN PHOSPHATASE PROTEINS, NUCLEIC ACID MOLECULES ENCODING HUMAN PHOSPHATASE PROTEINS, AND USES THEREOF

RELATED APPLICATIONS

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The present application is a Continuation-In-Part of U.S. Serial No. 09/715,177, filed November 20, 2000 and Continuation-In-Part of U.S. Serial No. 09/761,640.

FIELD OF THE INVENTION

The present invention is in the field of phosphatase proteins that are related to the mitogen-activated protein (MAP) kinase phosphatase subfamily, recombinant DNA molecules and protein production. The present invention provides novel phosphatase peptides and proteins and nucleic acid molecules encoding such peptide and protein molecules, all of which are useful in the development of human therapeutics and diagnostic compositions and methods. Specifically, the present invention provides three novel MAP kinase phosphatase splice forms.

BACKGROUND OF THE INVENTION

Phosphatase proteins, particularly members of the MAP kinase phosphatase subfamily, are a major target for drug action and development. Accordingly, it is valuable to the field of pharmaceutical development to identify and characterize previously unknown members of this subfamily of phosphatase proteins. The present invention advances the state of the art by providing a previously unidentified human phosphatase proteins that have homology to members of the MAP kinase phosphatase subfamily.

Protein Phosphatases

Cellular signal transduction is a fundamental mechanism whereby external stimuli that regulate diverse cellular processes are relayed to the interior of cells. The biochemical pathways through which signals are transmitted within cells comprise a circuitry of directly or functionally connected interactive proteins. One of the key biochemical mechanisms of signal transduction involves the reversible phosphorylation of certain residues on proteins. The phosphorylation state of a protein may affect its conformation and/or enzymic activity as well as its cellular location. The phosphorylation state of a protein is modified through the reciprocal actions of protein phosphatases (PKs) and protein phosphatases (PPs) at various specific amino acid

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residues.

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Protein phosphorylation is the ubiquitous strategy used to control the activities of eukaryotic cells. It is estimated that 10% of the proteins active in a typical mammalian cell are phosphorylated. The high-energy phosphate that confers activation and is transferred from adenosine triphosphate molecules to protein-by-protein phosphatases is subsequently removed from the protein-by-protein phosphatases. In this way, the phosphatases control most cellular signaling events that regulate cell growth and differentiation, cell-to-cell contacts, the cell cycle, and oncogenesis.

The protein phosphorylation/dephosphorylation cycle is one of the major regulatory mechanisms employed by eukaryotic cells to control cellular activities. It is estimated that more than 10% of the active proteins in a typical mammalian cell are phosphorylated. During protein phosphorylation/dephosphorylation, phosphate groups are transferred from adenosine triphosphate molecules to protein-by-protein phosphatases and are removed from the protein-by-protein phosphatases.

Protein phosphatases function in cellular signaling events that regulate cell growth and differentiation, cell-to-cell contacts, the cell cycle, and oncogenesis. Three protein phosphatase families have been identified as evolutionarily distinct. These include the serine/threonine phosphatases, the protein tyrosine phosphatases, and the acid/alkaline phosphatases (Carbonneau H. and Tonks N. K. (1992) Annu. Rev. Cell Biol. 8:463-93).

The serine/threonine phosphatases are either cytosolic or associated with a receptor. On the basis of their sensitivity to two thermostable proteins, inhibitors 1 and 2, and their divalent cation requirements, the serine/threonine phosphatases can be separated into four distinct groups, PP-I, PP-IIA, PP-IIB, and PP-IIC.

PP-I dephosphorylates many of the proteins phosphorylated by cylic AMP-dependent protein phosphatase and is therefore an important regulator of many cyclic AMP mediated, hormone responses in cells. PP-IIA has broad specificity for control of cell cycle, growth and proliferation, and DNA replication and is the main phosphatase responsible for reversing the phosphorylations of serine/threonine phosphatases. PP-IIB, or calcineurin (Cn), is a Ca.sup.+2 - activated phosphatase; it is involved in the regulation of such diverse cellular functions as ion channel regulation, neuronal transmission, gene transcription, muscle glycogen metabolism, and lymphocyte activation.

PP-IIC is a Mg.sup.++ -dependent phosphatase which participates in a wide variety of functions including regulating cyclic AMP-activated protein-phosphatase activity, Ca.sup.++ -

dependent signal transduction, tRNA splicing, and signal transmission related to heat shock responses. PP-IIC is a monomeric protein with a molecular mass of about 40-45 kDa. One alpha. and several .beta. isoforms of PP-IIC have been identified (Wenk, J. et al. (1992) FEBS Lett. 297: 135-138; Terasawa, T. et al. (1993) Arch. Biochem. Biophys. 307: 342-349; and Kato, S. et al. (1995) Arch. Biochem. Biophys. 318: 387-393).

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The levels of protein phosphorylation required for normal cell growth and differentiation at any time are achieved through the coordinated action of PKs and PPS. Depending on the cellular context, these two types of enzymes may either antagonize or cooperate with each other during signal transduction. An imbalance between these enzymes may impair normal cell functions leading to metabolic disorders and cellular transformation.

For example, insulin binding to the insulin receptor, which is a PTK, triggers a variety of metabolic and growth promoting effects such as glucose transport, biosynthesis of glycogen and fats, DNA synthesis, cell division and differentiation. Diabetes mellitus, which is characterized by insufficient or a lack of insulin signal transduction, can be caused by any abnormality at any step along the insulin signaling pathway. (Olefsky, 1988, in "Cecil Textbook of Medicine," 18th Ed., 2:1360-81).

It is also well known, for example, that the overexpression of PTKs, such as HER2, can play a decisive role in the development of cancer (Slamon et al., 1987, Science 235:77-82) and that antibodies capable of blocking the activity of this enzyme can abrogate tumor growth (Drebin et al., 1988, Oncogene 2:387-394). Blocking the signal transduction capability of tyrosine phosphatases such as Flk-1 and the PDGF receptor have been shown to block tumor growth in animal models (Millauer et al., 1994, Nature 367:577; Ueno et al., Science, 252:844-848).

Relatively less is known with respect to the direct role of phosphatases in signal transduction; PPs may play a role in human diseases. For example, ectopic expression of RPTP.alpha. produces a transformed phenotype in embryonic fibroblasts (Zheng et al., Nature 359:336-339), and overexpression of RPTP.alpha. in embryonal carcinoma cells causes the cells to differentiate into a cell type with neuronal phenotype (den Hertog et al., EMBO J 12:3789-3798). The gene for human RPTP.gamma. has been localized to chromosome 3p21 which is a segment frequently altered in renal and small lung carcinoma. Mutations may occur in the extracellular segment of RPTP.gamma. which renders a RPTP that no longer respond to external signals (LaForgia et al., Wary et al., 1993, Cancer Res 52:478-482). Mutations in the gene encoding PTP1C (also known as HCP, SHP) are the cause of the moth-eaten phenotype in mice

that suffer severe immunodeficiency, and systemic autoimmune disease accompanied by hyperproliferation of macrophages (Schultz et al., 1993, Cell 73:1445-1454). PTP1D (also known as Syp or PTP2C) has been shown to bind through SH2 domains to sites of phosphorylation in PDGFR, EGFR and insulin receptor substrate 1 (IRS-1). Reducing the activity of PTP1D by microinjection of anti-PTP1D antibody has been shown to block insulin or EGF-induced mitogenesis (Xiao et al., 1994, J Biol Chem 269:21244-21248).

MAP Kinase Phosphatases

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The present invention provides three novel alternative splice forms of mitogen-activated protein (MAP) kinase phosphatase. The alternative splice forms are herein referred to as splice forms 1, 2, and 3. Specifically, as indicated in Figure 3, splice form 2 includes exons 13 and 14, which are absent in splice forms 1 and 3, and splice form 3 is missing exon 7, which is present in splice forms 1 and 2. cDNA clones from all three isoforms have been mapped to the same region of human chromsome 11. Splice form 1 has been previously disclosed by applicant in U.S. application 09/715,177, filed November 20, 2000.

MAP kinase phosphatases are dual-specificity protein phosphatases involved in numerous critical biological processes. In *Drosophila*, MAP kinase phosphatases have been found to be essential for viability. Furthermore, loss-of-function mutations cause kinked and/or branched bristles and wing hairs. In plants, MAP kinase phosphatases play a critical role in responses to stress and pathogens (Gupta *et al.*, *Plant J* 1998 Dec;16(5):581-9). MAP kinase phosphatases play a critical role in neuronal survival and neuronal cell death following injury and degenerative stimuli (Winter *et al.*, *Brain Res* 1998 Aug 10;801(1-2):198-205). MAP kinase phosphatase may also play a role in diabetes and other insulin-related conditions; insulin regulates map kinase phosphatase-1 (MKP-1) and it has been suggested that MKP-1 acts as a negative regulator of insulin signaling (Kusari *et al.*, *Mol Endocrinol* 1997 Sep;11(10):1532-43). Furthermore, MAP kinase phosphatases may be critical for the negative regulation of cellular proliferation (Emslie *et al.*, *Hum Genet* 1994 May;93(5):513-6) and therefore, novel human MAP kinase phosphatase genes are useful as candidate tumor-suppressor genes. Additionally, MAP kinase phosphatases may specifically be involved in pancreatic cancer (Furukawa *et al.*, *Cytogenet Cell Genet* 1998;82(3-4):156-9).

For a further review of MAP kinase phosphatases, see Scimeca et al., Oncogene 1997 Aug 7;15(6):717-25. See Wang et al., Genomics 57 (2), 310-315 (1999) for supporting information on alternative splice variants.

The discovery of new human protein phosphatases and the polynucleotides encoding them satisfies a need in the art by providing new compositions that are useful in the diagnosis, prevention and treatment of biological processes associated with abnormal or unwanted protein phosphorylation.

SUMMARY OF THE INVENTION

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The present invention is based in part on the identification of amino acid sequences of human phosphatase peptides and proteins that are related to the MAP kinase phosphatase subfamily, as well as allelic variants and other mammalian orthologs thereof. Specifically, the present invention provides three novel MAP kinase phosphatase splice forms. These unique peptide sequences, and nucleic acid sequences that encode these peptides, can be used as models for the development of human therapeutic targets, aid in the identification of therapeutic proteins, and serve as targets for the development of human therapeutic agents that modulate phosphatase activity in cells and tissues that express the phosphatase. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain.

DESCRIPTION OF THE FIGURE SHEETS

FIGURE 1 provides the nucleotide sequence of three cDNA molecules that encode each of the phosphatase splice forms of the present invention. (splice form 1 = SEQ ID NO:1, splice form 2 = SEQ ID NO:4, splice form 3 = SEQ ID NO:6) In addition, structure and functional information is provided, such as ATG start, stop and tissue distribution, where available, that allows one to readily determine specific uses of inventions based on this molecular sequence. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain.

FIGURE 2 provides the predicted amino acid sequence of the three phosphatase splice forms of the present invention. (splice form 1 = SEQ ID NO:2, splice form 2 = SEQ ID NO:5, splice form 3 = SEQ ID NO:7) In addition structure and functional information such as protein family, function, and modification sites is provided where available, allowing one to readily determine specific uses of inventions based on this molecular sequence.

FIGURE 3 provides genomic sequences that span the gene encoding the three phosphatase splice forms of the present invention. (SEQ ID NO:3) In addition structure and functional information, such as intron/exon structure, promoter location, etc., is provided where available, allowing one to readily determine specific uses of inventions based on this molecular sequence. Figure 3 also indicates that the map position of the genomic sequence encoding the three splice forms is human chromosome 11. As illustrated in Figure 3, the following SNP variations were identified: G577A, G1451A, and G2641A. Figure 3 further provides a gene structure model and multiple alignments of the cDNA and peptide sequences of the three splice forms to illustrate the structure/sequence variations. Specifically, as indicated in Figure 3, splice form 2 includes exons 13 and 14, which are absent in splice forms 1 and 3, and splice form 3 is missing exon 7, which is present in splice forms 1 and 2.

DETAILED DESCRIPTION OF THE INVENTION

General Description

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The present invention is based on the sequencing of the human genome. During the sequencing and assembly of the human genome, analysis of the sequence information revealed previously unidentified fragments of the human genome that encode peptides that share structural and/or sequence homology to protein/peptide/domains identified and characterized within the art as being a phosphatase protein or part of a phosphatase protein and are related to the MAP kinase phosphatase subfamily. Utilizing these sequences, additional genomic sequences were assembled and transcript and/or cDNA sequences were isolated and characterized. Based on this analysis, the present invention provides amino acid sequences of three novel human MAP kinase phosphatase splice forms, nucleic acid sequences in the form of cDNA sequences and genomic sequences that encode these phosphatase splice forms, nucleic acid variation (allelic information), tissue distribution of expression, and information about the closest art known protein/peptide/domain that has structural or sequence homology to the phosphatase proteins of the present invention.

In addition to being previously unknown, the peptides that are provided in the present invention are selected based on their ability to be used for the development of commercially important products and services. Specifically, the present peptides are selected based on homology and/or structural relatedness to known phosphatase proteins of the MAP kinase phosphatase subfamily and the expression pattern observed. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. The art has clearly established the commercial importance of members of this family of proteins and proteins that have expression patterns similar to that of the present gene. Some of the more specific features of the peptides of the present invention, and the uses thereof, are described herein, particularly in the Background of the Invention and in the annotation provided in the Figures, and/or are known within the art for each of the known MAP kinase phosphatase family or subfamily of phosphatase proteins.

Specific Embodiments

Peptide Molecules

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The present invention provides nucleic acid sequences that encode protein molecules that have been identified as being members of the phosphatase family of proteins and are related to the MAP kinase phosphatase subfamily (protein sequences are provided in Figure 2, transcript/cDNA sequences are provided in Figure 1 and genomic sequences are provided in Figure 3). Specifically, the present invention provides three novel MAP kinase phosphatase splice forms. The peptide sequences provided in Figure 2, as well as the obvious variants described herein, particularly allelic variants as identified herein and using the information in Figure 3, will be referred herein as the phosphatase peptides of the present invention, phosphatase peptides, phosphatase splice forms, or peptides/proteins/splice forms of the present invention.

The present invention provides isolated peptide and protein molecules that consist of, consist essentially of, or comprise the amino acid sequences of the phosphatase peptides disclosed in the Figure 2, (encoded by the nucleic acid molecule shown in Figure 1, transcript/cDNA or Figure 3, genomic sequence), as well as all obvious variants of these

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peptides that are within the art to make and use. Some of these variants are described in detail below.

As used herein, a peptide is said to be "isolated" or "purified" when it is substantially free of cellular material or free of chemical precursors or other chemicals. The peptides of the present invention can be purified to homogeneity or other degrees of purity. The level of purification will be based on the intended use. The critical feature is that the preparation allows for the desired function of the peptide, even if in the presence of considerable amounts of other components (the features of an isolated nucleic acid molecule is discussed below).

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In some uses, "substantially free of cellular material" includes preparations of the peptide having less than about 30% (by dry weight) other proteins (i.e., contaminating protein), less than about 20% other proteins, less than about 10% other proteins, or less than about 5% other proteins. When the peptide is recombinantly produced, it can also be substantially free of culture medium, i.e., culture medium represents less than about 20% of the volume of the protein preparation.

The language "substantially free of chemical precursors or other chemicals" includes preparations of the peptide in which it is separated from chemical precursors or other chemicals that are involved in its synthesis. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of the phosphatase peptide having less than about 30% (by dry weight) chemical precursors or other chemicals, less than about 20% chemical precursors or other chemicals, less than about 10% chemical precursors or other chemicals, or less than about 5% chemical precursors or other chemicals.

The isolated phosphatase peptide can be purified from cells that naturally express it, purified from cells that have been altered to express it (recombinant), or synthesized using known protein synthesis methods. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. For example, a nucleic acid molecule encoding the phosphatase peptide is cloned into an expression vector, the expression vector introduced into a host cell and the protein expressed in the host cell. The protein can then be isolated from the cells by an appropriate purification scheme using standard protein purification techniques. Many of these techniques are described in detail below.

Accordingly, the present invention provides proteins that consist of the amino acid sequences provided in Figure 2 (SEQ ID NOS:2,5,7), for example, proteins encoded by the cDNA

nucleic acid sequences shown in Figure 1 (SEQ ID NOS:1,4,6) and the genomic sequences provided in Figure 3 (SEQ ID NO:3). The amino acid sequence of such a protein is provided in Figure 2. A protein consists of an amino acid sequence when the amino acid sequence is the final amino acid sequence of the protein.

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The present invention further provides proteins that consist essentially of the amino acid sequences provided in Figure 2 (SEQ ID NOS:2,5,7), for example, proteins encoded by the cDNA nucleic acid sequences shown in Figure 1 (SEQ ID NOS:1,4,6) and the genomic sequences provided in Figure 3 (SEQ ID NO:3). A protein consists essentially of an amino acid sequence when such an amino acid sequence is present with only a few additional amino acid residues, for example from about 1 to about 100 or so additional residues, typically from 1 to about 20 additional residues in the final protein.

The present invention further provides proteins that comprise the amino acid sequences provided in Figure 2 (SEQ ID NOS:2,5,7), for example, proteins encoded by the cDNA nucleic acid sequences shown in Figure 1 (SEQ ID NOS:1,4,6) and the genomic sequences provided in Figure 3 (SEQ ID NO:3). A protein comprises an amino acid sequence when the amino acid sequence is at least part of the final amino acid sequence of the protein. In such a fashion, the protein can be only the peptide or have additional amino acid molecules, such as amino acid residues (contiguous encoded sequence) that are naturally associated with it or heterologous amino acid residues/peptide sequences. Such a protein can have a few additional amino acid residues or can comprise several hundred or more additional amino acids. The preferred classes of proteins that are comprised of the phosphatase peptides of the present invention are the naturally occurring mature proteins. A brief description of how various types of these proteins can be made/isolated is provided below.

The phosphatase peptides of the present invention can be attached to heterologous sequences to form chimeric or fusion proteins. Such chimeric and fusion proteins comprise a phosphatase peptide operatively linked to a heterologous protein having an amino acid sequence not substantially homologous to the phosphatase peptide. "Operatively linked" indicates that the phosphatase peptide and the heterologous protein are fused in-frame. The heterologous protein can be fused to the N-terminus or C-terminus of the phosphatase peptide.

In some uses, the fusion protein does not affect the activity of the phosphatase peptide per se. For example, the fusion protein can include, but is not limited to, enzymatic fusion proteins, for example beta-galactosidase fusions, yeast two-hybrid GAL fusions, poly-His fusions, MYC-tagged, HI-tagged and Ig fusions. Such fusion proteins, particularly poly-His fusions, can facilitate the purification of recombinant phosphatase peptide. In certain host cells (e.g., mammalian host cells),

expression and/or secretion of a protein can be increased by using a heterologous signal sequence.

A chimeric or fusion protein can be produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different protein sequences are ligated together inframe in accordance with conventional techniques. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and re-amplified to generate a chimeric gene sequence (see Ausubel et al., Current Protocols in Molecular Biology, 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (e.g., a GST protein). A phosphatase peptide-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the phosphatase peptide.

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As mentioned above, the present invention also provides and enables obvious variants of the amino acid sequence of the proteins of the present invention, such as naturally occurring mature forms of the peptide, allelic/sequence variants of the peptides, non-naturally occurring recombinantly derived variants of the peptides, and orthologs and paralogs of the peptides. Such variants can readily be generated using art-known techniques in the fields of recombinant nucleic acid technology and protein biochemistry. It is understood, however, that variants exclude any amino acid sequences disclosed prior to the invention.

Such variants can readily be identified/made using molecular techniques and the sequence information disclosed herein. Further, such variants can readily be distinguished from other peptides based on sequence and/or structural homology to the phosphatase peptides of the present invention. The degree of homology/identity present will be based primarily on whether the peptide is a functional variant or non-functional variant, the amount of divergence present in the paralog family and the evolutionary distance between the orthologs.

To determine the percent identity of two amino acid sequences or two nucleic acid sequences, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in one or both of a first and a second amino acid or nucleic acid sequence for optimal alignment and non-homologous sequences can be disregarded for comparison purposes). In a preferred embodiment, at least 30%, 40%, 50%, 60%, 70%, 80%, or 90% or more of the length of a reference sequence is aligned for comparison purposes. The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide

as the corresponding position in the second sequence, then the molecules are identical at that position (as used herein amino acid or nucleic acid "identity" is equivalent to amino acid or nucleic acid "homology"). The percent identity between the two sequences is a function of the number of identical positions shared by the sequences, taking into account the number of gaps, and the length of each gap, which need to be introduced for optimal alignment of the two sequences.

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The comparison of sequences and determination of percent identity and similarity between two sequences can be accomplished using a mathematical algorithm. (Computational Molecular Biology, Lesk, A.M., ed., Oxford University Press, New York, 1988; Biocomputing: Informatics and Genome Projects, Smith, D.W., ed., Academic Press, New York, 1993; Computer Analysis of Sequence Data, Part 1, Griffin, A.M., and Griffin, H.G., eds., Humana Press, New Jersey, 1994; Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press, 1987; and Sequence Analysis Primer, Gribskov, M. and Devereux, J., eds., M Stockton Press, New York, 1991). In a preferred embodiment, the percent identity between two amino acid sequences is determined using the Needleman and Wunsch (J. Mol. Biol. (48):444-453 (1970)) algorithm which has been incorporated into the GAP program in the GCG software package (available at http://www.gcg.com), using either a Blossom 62 matrix or a PAM250 matrix, and a gap weight of 16, 14, 12, 10, 8, 6, or 4 and a length weight of 1, 2, 3, 4, 5, or 6. In yet another preferred embodiment, the percent identity between two nucleotide sequences is determined using the GAP program in the GCG software package (Devereux, J., et al., Nucleic Acids Res. 12(1):387 (1984)) (available at http://www.gcg.com), using a NWSgapdna.CMP matrix and a gap weight of 40, 50, 60, 70, or 80 and a length weight of 1, 2, 3, 4, 5, or 6. In another embodiment, the percent identity between two amino acid or nucleotide sequences is determined using the algorithm of E. Myers and W. Miller (CABIOS, 4:11-17 (1989)) which has been incorporated into the ALIGN program (version 2.0), using a PAM120 weight residue table, a gap length penalty of 12 and a gap penalty of 4.

The nucleic acid and protein sequences of the present invention can further be used as a "query sequence" to perform a search against sequence databases to, for example, identify other family members or related sequences. Such searches can be performed using the NBLAST and XBLAST programs (version 2.0) of Altschul, et al. (J. Mol. Biol. 215:403-10 (1990)). BLAST nucleotide searches can be performed with the NBLAST program, score = 100, wordlength = 12 to obtain nucleotide sequences homologous to the nucleic acid molecules of the invention. BLAST protein searches can be performed with the XBLAST program, score = 50, wordlength =

3 to obtain amino acid sequences homologous to the proteins of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST can be utilized as described in Altschul et al. (Nucleic Acids Res. 25(17):3389-3402 (1997)). When utilizing BLAST and gapped BLAST programs, the default parameters of the respective programs (e.g., XBLAST and NBLAST) can be used.

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Full-length pre-processed forms, as well as mature processed forms, of proteins that comprise one of the peptides of the present invention can readily be identified as having complete sequence identity to one of the phosphatase peptides of the present invention as well as being encoded by the same genetic locus as the phosphatase peptide provided herein. The gene encoding the novel phosphatase proteins of the present invention is located on public BAC AP001885, which is known to be mapped to chromosome 11 (as indicated in Figure 3).

Allelic variants of a phosphatase peptide can readily be identified as being a human protein having a high degree (significant) of sequence homology/identity to at least a portion of the phosphatase peptide as well as being encoded by the same genetic locus as the phosphatase peptide provided herein. Genetic locus can readily be determined based on the genomic information provided in Figure 3, such as the genomic sequence mapped to the reference human. The gene encoding the novel phosphatase proteins of the present invention is located on public BAC AP001885, which is known to be mapped to chromosome 11 (as indicated in Figure 3). As used herein, two proteins (or a region of the proteins) have significant homology when the amino acid sequences are typically at least about 70-80%, 80-90%, and more typically at least about 90-95% or more homologous. A significantly homologous amino acid sequence, according to the present invention, will be encoded by a nucleic acid sequence that will hybridize to a phosphatase peptide encoding nucleic acid molecule under stringent conditions as more fully described below.

Figure 3 provides information on SNPs that have been found in the gene encoding the phosphatase proteins of the present invention. The following SNPs were identified: G577A, G1451A, and G2641A. G577A and G1451A are non-synonymous coding SNPs. Changes in the amino acid sequence caused by these SNPs is indicated in Figure 3 and can readily be determined using the universal genetic code and the protein sequence provided in Figure 2 as a reference. G2641A is 3' of the ORF and may affect control/regulatory elements.

Paralogs of a phosphatase peptide can readily be identified as having some degree of significant sequence homology/identity to at least a portion of the phosphatase peptide, as being encoded by a gene from humans, and as having similar activity or function. Two proteins will

typically be considered paralogs when the amino acid sequences are typically at least about 60% or greater, and more typically at least about 70% or greater homology through a given region or domain. Such paralogs will be encoded by a nucleic acid sequence that will hybridize to a phosphatase peptide encoding nucleic acid molecule under moderate to stringent conditions as more fully described below.

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Orthologs of a phosphatase peptide can readily be identified as having some degree of significant sequence homology/identity to at least a portion of the phosphatase peptide as well as being encoded by a gene from another organism. Preferred orthologs will be isolated from mammals, preferably primates, for the development of human therapeutic targets and agents. Such orthologs will be encoded by a nucleic acid sequence that will hybridize to a phosphatase peptide encoding nucleic acid molecule under moderate to stringent conditions, as more fully described below, depending on the degree of relatedness of the two organisms yielding the proteins.

Non-naturally occurring variants of the phosphatase peptides of the present invention can readily be generated using recombinant techniques. Such variants include, but are not limited to deletions, additions and substitutions in the amino acid sequence of the phosphatase peptide. For example, one class of substitutions are conserved amino acid substitution. Such substitutions are those that substitute a given amino acid in a phosphatase peptide by another amino acid of like characteristics. Typically seen as conservative substitutions are the replacements, one for another, among the aliphatic amino acids Ala, Val, Leu, and Ile; interchange of the hydroxyl residues Ser and Thr; exchange of the acidic residues Asp and Glu; substitution between the amide residues Asn and Gln; exchange of the basic residues Lys and Arg; and replacements among the aromatic residues Phe and Tyr. Guidance concerning which amino acid changes are likely to be phenotypically silent are found in Bowie et al., Science 247:1306-1310 (1990).

Variant phosphatase peptides can be fully functional or can lack function in one or more activities, e.g. ability to bind substrate, ability to dephosphorylate substrate, ability to mediate signaling, etc. Fully functional variants typically contain only conservative variation or variation in non-critical residues or in non-critical regions. Figure 2 provides the result of protein analysis and can be used to identify critical domains/regions. Functional variants can also contain substitution of similar amino acids that result in no change or an insignificant change in function. Alternatively, such substitutions may positively or negatively affect function to some degree.

Non-functional variants typically contain one or more non-conservative amino acid substitutions, deletions, insertions, inversions, or truncation or a substitution, insertion, inversion, or deletion in a critical residue or critical region.

Amino acids that are essential for function can be identified by methods known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham et al., Science 244:1081-1085 (1989)), particularly using the results provided in Figure 2. The latter procedure introduces single alanine mutations at every residue in the molecule. The resulting mutant molecules are then tested for biological activity such as phosphatase activity or in assays such as an in vitro proliferative activity. Sites that are critical for binding partner/substrate binding can also be determined by structural analysis such as crystallization, nuclear magnetic resonance or photoaffinity labeling (Smith et al., J. Mol. Biol. 224:899-904 (1992); de Vos et al. Science 255:306-312 (1992)).

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The present invention further provides fragments of the phosphatase peptides, in addition to proteins and peptides that comprise and consist of such fragments, particularly those comprising the residues identified in Figure 2. The fragments to which the invention pertains, however, are not to be construed as encompassing fragments that may be disclosed publicly prior to the present invention.

As used herein, a fragment comprises at least 8, 10, 12, 14, 16, or more contiguous amino acid residues from a phosphatase peptide. Such fragments can be chosen based on the ability to retain one or more of the biological activities of the phosphatase peptide or could be chosen for the ability to perform a function, e.g. bind a substrate or act as an immunogen. Particularly important fragments are biologically active fragments, peptides that are, for example, about 8 or more amino acids in length. Such fragments will typically comprise a domain or motif of the phosphatase peptide, e.g., active site, a transmembrane domain or a substrate-binding domain. Further, possible fragments include, but are not limited to, domain or motif containing fragments, soluble peptide fragments, and fragments containing immunogenic structures. Predicted domains and functional sites are readily identifiable by computer programs well known and readily available to those of skill in the art (e.g., PROSITE analysis). The results of one such analysis are provided in Figure 2.

Polypeptides often contain amino acids other than the 20 amino acids commonly referred to as the 20 naturally occurring amino acids. Further, many amino acids, including the terminal amino acids, may be modified by natural processes, such as processing and other post-translational modifications, or by chemical modification techniques well known in the art. Common modifications that occur naturally in phosphatase peptides are described in basic texts, detailed monographs, and the research literature, and they are well known to those of skill in the art (some of these features are identified in Figure 2).

Known modifications include, but are not limited to, acetylation, acylation, ADP-

ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of phosphotidylinositol, cross-linking, cyclization, disulfide bond formation, demethylation, formation of covalent crosslinks, formation of cystine, formation of pyroglutamate, formylation, gamma carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, transfer-RNA mediated addition of amino acids to proteins such as arginylation, and ubiquitination.

Such modifications are well known to those of skill in the art and have been described in great detail in the scientific literature. Several particularly common modifications, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation, for instance, are described in most basic texts, such as *Proteins - Structure and Molecular Properties*, 2nd Ed., T.E. Creighton, W. H. Freeman and Company, New York (1993). Many detailed reviews are available on this subject, such as by Wold, F., *Posttranslational Covalent Modification of Proteins*, B.C. Johnson, Ed., Academic Press, New York 1-12 (1983); Seifter *et al.* (*Meth. Enzymol. 182*: 626-646 (1990)) and Rattan *et al.* (*Ann. N.Y. Acad. Sci. 663*:48-62 (1992)).

Accordingly, the phosphatase peptides of the present invention also encompass derivatives or analogs in which a substituted amino acid residue is not one encoded by the genetic code, in which a substituent group is included, in which the mature phosphatase peptide is fused with another compound, such as a compound to increase the half-life of the phosphatase peptide, or in which the additional amino acids are fused to the mature phosphatase peptide, such as a leader or secretory sequence or a sequence for purification of the mature phosphatase peptide or a pro-protein sequence.

25 <u>Protein/Peptide Uses</u>

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The proteins of the present invention can be used in substantial and specific assays related to the functional information provided in the Figures; to raise antibodies or to elicit another immune response; as a reagent (including the labeled reagent) in assays designed to quantitatively determine levels of the protein (or its binding partner or ligand) in biological fluids; and as markers for tissues in which the corresponding protein is preferentially expressed (either constitutively or at a particular stage of tissue differentiation or development or in a disease state). Where the protein binds or potentially binds to another protein or ligand (such as,

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for example, in a phosphatase-effector protein interaction or phosphatase-ligand interaction), the protein can be used to identify the binding partner/ligand so as to develop a system to identify inhibitors of the binding interaction. Any or all of these uses are capable of being developed into reagent grade or kit format for commercialization as commercial products.

Methods for performing the uses listed above are well known to those skilled in the art. References disclosing such methods include "Molecular Cloning: A Laboratory Manual", 2d ed., Cold Spring Harbor Laboratory Press, Sambrook, J., E. F. Fritsch and T. Maniatis eds., 1989, and "Methods in Enzymology: Guide to Molecular Cloning Techniques", Academic Press, Berger, S. L. and A. R. Kimmel eds., 1987.

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The potential uses of the peptides of the present invention are based primarily on the source of the protein as well as the class/action of the protein. For example, phosphatases isolated from humans and their human/mammalian orthologs serve as targets for identifying agents for use in mammalian therapeutic applications, e.g. a human drug, particularly in modulating a biological or pathological response in a cell or tissue that expresses the phosphatase. Experimental data as provided in Figure 1 indicates that splice form 1 of the phosphatase protein of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Specifically, a virtual northern blot shows expression in pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, and ovary tumor tissue. In addition, PCR-based tissue screening panels indicate expression in the brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. A large percentage of pharmaceutical agents are being developed that modulate the activity of phosphatase proteins, particularly members of the MAP kinase phosphatase subfamily (see Background of the Invention). The structural and functional information provided in the Background and Figures provide specific and substantial uses for the molecules of the present invention, particularly in combination with the expression information provided in Figure 1. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is

expressed in fetal brain. Such uses can readily be determined using the information provided herein, that which is known in the art, and routine experimentation.

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The proteins of the present invention (including variants and fragments that may have been disclosed prior to the present invention) are useful for biological assays related to phosphatases that are related to members of the MAP kinase phosphatase subfamily. Such assays involve any of the known phosphatase functions or activities or properties useful for diagnosis and treatment of phosphatase-related conditions that are specific for the subfamily of phosphatases that the one of the present invention belongs to, particularly in cells and tissues that express the phosphatase. Experimental data as provided in Figure 1 indicates that splice form 1 of the phosphatase protein of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Specifically, a virtual northern blot shows expression in pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, and ovary tumor tissue. In addition, PCR-based tissue screening panels indicate expression in the brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid.

The proteins of the present invention are also useful in drug screening assays, in cell-based or cell-free systems. Cell-based systems can be native, i.e., cells that normally express the phosphatase, as a biopsy or expanded in cell culture. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. In an alternate embodiment, cell-based assays involve recombinant host cells expressing the phosphatase protein.

The polypeptides can be used to identify compounds that modulate phosphatase activity of the protein in its natural state or an altered form that causes a specific disease or pathology associated with the phosphatase. Both the phosphatases of the present invention and appropriate variants and fragments can be used in high-throughput screens to assay candidate compounds for the ability to bind to the phosphatase. These compounds can be further screened against a functional phosphatase to determine the effect of the compound on the phosphatase activity.

Further, these compounds can be tested in animal or invertebrate systems to determine activity/effectiveness. Compounds can be identified that activate (agonist) or inactivate (antagonist) the phosphatase to a desired degree.

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Further, the proteins of the present invention can be used to screen a compound for the ability to stimulate or inhibit interaction between the phosphatase protein and a molecule that normally interacts with the phosphatase protein, e.g. a substrate or a component of the signal pathway that the phosphatase protein normally interacts (for example, another phosphatase). Such assays typically include the steps of combining the phosphatase protein with a candidate compound under conditions that allow the phosphatase protein, or fragment, to interact with the target molecule, and to detect the formation of a complex between the protein and the target or to detect the biochemical consequence of the interaction with the phosphatase protein and the target, such as any of the associated effects of signal transduction such as protein phosphorylation, cAMP turnover, and adenylate cyclase activation, etc.

Candidate compounds include, for example, 1) peptides such as soluble peptides, including Ig-tailed fusion peptides and members of random peptide libraries (see, e.g., Lam et al., Nature 354:82-84 (1991); Houghten et al., Nature 354:84-86 (1991)) and combinatorial chemistry-derived molecular libraries made of D- and/or L- configuration amino acids; 2) phosphopeptides (e.g., members of random and partially degenerate, directed phosphopeptide libraries, see, e.g., Songyang et al., Cell 72:767-778 (1993)); 3) antibodies (e.g., polyclonal, monoclonal, humanized, anti-idiotypic, chimeric, and single chain antibodies as well as Fab, F(ab')₂, Fab expression library fragments, and epitope-binding fragments of antibodies); and 4) small organic and inorganic molecules (e.g., molecules obtained from combinatorial and natural product libraries).

One candidate compound is a soluble fragment of the receptor that competes for substrate binding. Other candidate compounds include mutant phosphatases or appropriate fragments containing mutations that affect phosphatase function and thus compete for substrate. Accordingly, a fragment that competes for substrate, for example with a higher affinity, or a fragment that binds substrate but does not allow release, is encompassed by the invention.

The invention further includes other end point assays to identify compounds that modulate (stimulate or inhibit) phosphatase activity. The assays typically involve an assay of events in the signal transduction pathway that indicate phosphatase activity. Thus, the dephosphorylation of a substrate, activation of a protein, a change in the expression of genes that are up- or down-regulated in response to the phosphatase protein dependent signal cascade can be assayed.

Any of the biological or biochemical functions mediated by the phosphatase can be used as

an endpoint assay. These include all of the biochemical or biochemical/biological events described herein, in the references cited herein, incorporated by reference for these endpoint assay targets, and other functions known to those of ordinary skill in the art or that can be readily identified using the information provided in the Figures, particularly Figure 2. Specifically, a biological function of a cell or tissues that expresses the phosphatase can be assayed. Experimental data as provided in Figure 1 indicates that splice form 1 of the phosphatase protein of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Specifically, a virtual northern blot shows expression in pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, and ovary tumor tissue. In addition, PCR-based tissue screening panels indicate expression in the brain (including fetal), heart (including fetal), uterus, and thyroid.

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Binding and/or activating compounds can also be screened by using chimeric phosphatase proteins in which the amino terminal extracellular domain, or parts thereof, the entire transmembrane domain or subregions, such as any of the seven transmembrane segments or any of the intracellular or extracellular loops and the carboxy terminal intracellular domain, or parts thereof, can be replaced by heterologous domains or subregions. For example, a substrate-binding region can be used that interacts with a different substrate then that which is recognized by the native phosphatase. Accordingly, a different set of signal transduction components is available as an end-point assay for activation. This allows for assays to be performed in other than the specific host cell from which the phosphatase is derived.

The proteins of the present invention are also useful in competition binding assays in methods designed to discover compounds that interact with the phosphatase (e.g. binding partners and/or ligands). Thus, a compound is exposed to a phosphatase polypeptide under conditions that allow the compound to bind or to otherwise interact with the polypeptide. Soluble phosphatase polypeptide is also added to the mixture. If the test compound interacts with the soluble phosphatase polypeptide, it decreases the amount of complex formed or activity from the phosphatase target. This type of assay is particularly useful in cases in which compounds are sought that interact with specific regions of the phosphatase. Thus, the soluble polypeptide that competes with the target phosphatase region is designed to contain peptide sequences corresponding to the region of interest.

To perform cell free drug screening assays, it is sometimes desirable to immobilize either the phosphatase protein, or fragment, or its target molecule to facilitate separation of complexes from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay.

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Techniques for immobilizing proteins on matrices can be used in the drug screening assays. In one embodiment, a fusion protein can be provided which adds a domain that allows the protein to be bound to a matrix. For example, glutathione-S-transferase fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the cell lysates (e.g., 35S-labeled) and the candidate compound, and the mixture incubated under conditions conducive to complex formation (e.g., at physiological conditions for salt and pH). Following incubation, the beads are washed to remove any unbound label, and the matrix immobilized and radiolabel determined directly, or in the supernatant after the complexes are dissociated. Alternatively, the complexes can be dissociated from the matrix, separated by SDS-PAGE, and the level of phosphatase-binding protein found in the bead fraction quantitated from the gel using standard electrophoretic techniques. For example, either the polypeptide or its target molecule can be immobilized utilizing conjugation of biotin and streptavidin using techniques well known in the art. Alternatively, antibodies reactive with the protein but which do not interfere with binding of the protein to its target molecule can be derivatized to the wells of the plate, and the protein trapped in the wells by antibody conjugation. Preparations of a phosphatase-binding protein and a candidate compound are incubated in the phosphatase protein-presenting wells and the amount of complex trapped in the well can be quantitated. Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the phosphatase protein target molecule, or which are reactive with phosphatase protein and compete with the target molecule, as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the target molecule.

Agents that modulate one of the phosphatases of the present invention can be identified using one or more of the above assays, alone or in combination. It is generally preferable to use a cell-based or cell free system first and then confirm activity in an animal or other model system. Such model systems are well known in the art and can readily be employed in this context.

Modulators of phosphatase protein activity identified according to these drug screening assays can be used to treat a subject with a disorder mediated by the kinase pathway, by treating cells or tissues that express the phosphatase. Experimental data as provided in Figure 1 indicates

that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. These methods of treatment include the steps of administering a modulator of phosphatase activity in a pharmaceutical composition to a subject in need of such treatment, the modulator being identified as described herein.

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In yet another aspect of the invention, the phosphatase proteins can be used as "bait proteins" in a two-hybrid assay or three-hybrid assay (see, e.g., U.S. Patent No. 5,283,317; Zervos et al. (1993) Cell 72:223-232; Madura et al. (1993) J. Biol. Chem. 268:12046-12054; Bartel et al. (1993) Biotechniques 14:920-924; Iwabuchi et al. (1993) Oncogene 8:1693-1696; and Brent WO94/10300), to identify other proteins, which bind to or interact with the phosphatase and are involved in phosphatase activity. Such phosphatase-binding proteins are also likely to be involved in the propagation of signals by the phosphatase proteins or phosphatase targets as, for example, downstream elements of a kinase-mediated signaling pathway. Alternatively, such phosphatase-binding proteins are likely to be phosphatase inhibitors.

The two-hybrid system is based on the modular nature of most transcription factors, which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. In one construct, the gene that codes for a phosphatase protein is fused to a gene encoding the DNA binding domain of a known transcription factor (e.g., GAL-4). In the other construct, a DNA sequence, from a library of DNA sequences, that encodes an unidentified protein ("prey" or "sample") is fused to a gene that codes for the activation domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact, *in vivo*, forming a phosphatase-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (e.g., LacZ) which is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be detected and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene which encodes the protein which interacts with the phosphatase protein.

This invention further pertains to novel agents identified by the above-described screening assays. Accordingly, it is within the scope of this invention to further use an agent identified as described herein in an appropriate animal model. For example, an agent identified

as described herein (e.g., a phosphatase-modulating agent, an antisense phosphatase nucleic acid molecule, a phosphatase-specific antibody, or a phosphatase-binding partner) can be used in an animal or other model to determine the efficacy, toxicity, or side effects of treatment with such an agent. Alternatively, an agent identified as described herein can be used in an animal or other model to determine the mechanism of action of such an agent. Furthermore, this invention pertains to uses of novel agents identified by the above-described screening assays for treatments as described herein.

The phosphatase proteins of the present invention are also useful to provide a target for diagnosing a disease or predisposition to disease mediated by the peptide. Accordingly, the invention provides methods for detecting the presence, or levels of, the protein (or encoding mRNA) in a cell, tissue, or organism. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. The method involves contacting a biological sample with a compound capable of interacting with the phosphatase protein such that the interaction can be detected. Such an assay can be provided in a single detection format or a multi-detection format such as an antibody chip array.

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One agent for detecting a protein in a sample is an antibody capable of selectively binding to protein. A biological sample includes tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject.

The peptides of the present invention also provide targets for diagnosing active protein activity, disease, or predisposition to disease, in a patient having a variant peptide, particularly activities and conditions that are known for other members of the family of proteins to which the present one belongs. Thus, the peptide can be isolated from a biological sample and assayed for the presence of a genetic mutation that results in aberrant peptide. This includes amino acid substitution, deletion, insertion, rearrangement, (as the result of aberrant splicing events), and inappropriate post-translational modification. Analytic methods include altered electrophoretic mobility, altered tryptic peptide digest, altered phosphatase activity in cell-based or cell-free assay, alteration in substrate or antibody-binding pattern, altered isoelectric point, direct amino acid sequencing, and any other of the known assay techniques useful for detecting mutations in a protein. Such an assay can be provided in a single detection format or a multi-detection format such as an antibody chip array.

In vitro techniques for detection of peptide include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence using a detection reagent, such as an antibody or protein binding agent. Alternatively, the peptide can be detected *in vivo* in a subject by introducing into the subject a labeled anti-peptide antibody or other types of detection agent. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques. Particularly useful are methods that detect the allelic variant of a peptide expressed in a subject and methods which detect fragments of a peptide in a sample.

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The peptides are also useful in pharmacogenomic analysis. Pharmacogenomics deal with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See, e.g., Eichelbaum, M. (Clin. Exp. Pharmacol. Physiol. 23(10-11):983-985 (1996)), and Linder, M.W. (Clin. Chem. 43(2):254-266 (1997)). The clinical outcomes of these variations result in severe toxicity of therapeutic drugs in certain individuals or therapeutic failure of drugs in certain individuals as a result of individual variation in metabolism. Thus, the genotype of the individual can determine the way a therapeutic compound acts on the body or the way the body metabolizes the compound. Further, the activity of drug metabolizing enzymes effects both the intensity and duration of drug action. Thus, the pharmacogenomics of the individual permit the selection of effective compounds and effective dosages of such compounds for prophylactic or therapeutic treatment based on the individual's genotype. The discovery of genetic polymorphisms in some drug metabolizing enzymes has explained why some patients do not obtain the expected drug effects, show an exaggerated drug effect, or experience serious toxicity from standard drug dosages. Polymorphisms can be expressed in the phenotype of the extensive metabolizer and the phenotype of the poor metabolizer. Accordingly, genetic polymorphism may lead to allelic protein variants of the phosphatase protein in which one or more of the phosphatase functions in one population is different from those in another population. The peptides thus allow a target to ascertain a genetic predisposition that can affect treatment modality. Thus, in a ligandbased treatment, polymorphism may give rise to amino terminal extracellular domains and/or other substrate-binding regions that are more or less active in substrate binding, and phosphatase activation. Accordingly, substrate dosage would necessarily be modified to maximize the therapeutic effect within a given population containing a polymorphism. As an alternative to genotyping, specific polymorphic peptides could be identified.

The peptides are also useful for treating a disorder characterized by an absence of, inappropriate, or unwanted expression of the protein. Experimental data as provided in Figure 1

indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Accordingly, methods for treatment include the use of the phosphatase protein or fragments.

Antibodies

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The invention also provides antibodies that selectively bind to one of the peptides of the present invention, a protein comprising such a peptide, as well as variants and fragments thereof. As used herein, an antibody selectively binds a target peptide when it binds the target peptide and does not significantly bind to unrelated proteins. An antibody is still considered to selectively bind a peptide even if it also binds to other proteins that are not substantially homologous with the target peptide so long as such proteins share homology with a fragment or domain of the peptide target of the antibody. In this case, it would be understood that antibody binding to the peptide is still selective despite some degree of cross-reactivity.

As used herein, an antibody is defined in terms consistent with that recognized within the art: they are multi-subunit proteins produced by a mammalian organism in response to an antigen challenge. The antibodies of the present invention include polyclonal antibodies and monoclonal antibodies, as well as fragments of such antibodies, including, but not limited to, Fab or F(ab)₂, and Fv fragments.

Many methods are known for generating and/or identifying antibodies to a given target peptide. Several such methods are described by Harlow, Antibodies, Cold Spring Harbor Press, (1989).

In general, to generate antibodies, an isolated peptide is used as an immunogen and is administered to a mammalian organism, such as a rat, rabbit or mouse. The full-length protein, an antigenic peptide fragment or a fusion protein can be used. Particularly important fragments are those covering functional domains, such as the domains identified in Figure 2, and domain of sequence homology or divergence amongst the family, such as those that can readily be identified using protein alignment methods and as presented in the Figures.

Antibodies are preferably prepared from regions or discrete fragments of the phosphatase proteins. Antibodies can be prepared from any region of the peptide as described herein.

However, preferred regions will include those involved in function/activity and/or phosphatase/binding partner interaction. Figure 2 can be used to identify particularly important regions while sequence alignment can be used to identify conserved and unique sequence fragments.

An antigenic fragment will typically comprise at least 8 contiguous amino acid residues. The antigenic peptide can comprise, however, at least 10, 12, 14, 16 or more amino acid residues. Such fragments can be selected on a physical property, such as fragments correspond to regions that are located on the surface of the protein, e.g., hydrophilic regions or can be selected based on sequence uniqueness (see Figure 2).

Detection on an antibody of the present invention can be facilitated by coupling (i.e., physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, β-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ¹²⁵I, ¹³¹I, ³⁵S or ³H.

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Antibody Uses

The antibodies can be used to isolate one of the proteins of the present invention by standard techniques, such as affinity chromatography or immunoprecipitation. The antibodies can facilitate the purification of the natural protein from cells and recombinantly produced protein expressed in host cells. In addition, such antibodies are useful to detect the presence of one of the proteins of the present invention in cells or tissues to determine the pattern of expression of the protein among various tissues in an organism and over the course of normal development. Experimental data as provided in Figure 1 indicates that splice form 1 of the phosphatase protein of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Specifically, a virtual northern blot

shows expression in pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, and ovary tumor tissue. In addition, PCR-based tissue screening panels indicate expression in the brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Further, such antibodies can be used to detect protein *in situ*, *in vitro*, or in a cell lysate or supernatant in order to evaluate the abundance and pattern of expression. Also, such antibodies can be used to assess abnormal tissue distribution or abnormal expression during development or progression of a biological condition. Antibody detection of circulating fragments of the full length protein can be used to identify turnover.

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Further, the antibodies can be used to assess expression in disease states such as in active stages of the disease or in an individual with a predisposition toward disease related to the protein's function. When a disorder is caused by an inappropriate tissue distribution, developmental expression, level of expression of the protein, or expressed/processed form, the antibody can be prepared against the normal protein. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. If a disorder is characterized by a specific mutation in the protein, antibodies specific for this mutant protein can be used to assay for the presence of the specific mutant protein.

The antibodies can also be used to assess normal and aberrant subcellular localization of cells in the various tissues in an organism. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. The diagnostic uses can be applied, not only in genetic testing, but also in monitoring a treatment modality. Accordingly, where treatment is ultimately aimed at correcting expression level or the presence of aberrant sequence and aberrant tissue distribution or developmental expression, antibodies directed against the protein or relevant fragments can be used to monitor therapeutic efficacy.

Additionally, antibodies are useful in pharmacogenomic analysis. Thus, antibodies prepared against polymorphic proteins can be used to identify individuals that require modified treatment

modalities. The antibodies are also useful as diagnostic tools as an immunological marker for aberrant protein analyzed by electrophoretic mobility, isoelectric point, tryptic peptide digest, and other physical assays known to those in the art.

The antibodies are also useful for tissue typing. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Thus, where a specific protein has been correlated with expression in a specific tissue, antibodies that are specific for this protein can be used to identify a tissue type.

The antibodies are also useful for inhibiting protein function, for example, blocking the binding of the phosphatase peptide to a binding partner such as a substrate. These uses can also be applied in a therapeutic context in which treatment involves inhibiting the protein's function. An antibody can be used, for example, to block binding, thus modulating (agonizing or antagonizing) the peptides activity. Antibodies can be prepared against specific fragments containing sites required for function or against intact protein that is associated with a cell or cell membrane. See Figure 2 for structural information relating to the proteins of the present invention.

The invention also encompasses kits for using antibodies to detect the presence of a protein in a biological sample. The kit can comprise antibodies such as a labeled or labelable antibody and a compound or agent for detecting protein in a biological sample; means for determining the amount of protein in the sample; means for comparing the amount of protein in the sample with a standard; and instructions for use. Such a kit can be supplied to detect a single protein or epitope or can be configured to detect one of a multitude of epitopes, such as in an antibody detection array. Arrays are described in detail below for nuleic acid arrays and similar methods have been developed for antibody arrays.

Nucleic Acid Molecules

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The present invention further provides isolated nucleic acid molecules that encode a phosphatase peptide or protein of the present invention (cDNA, transcript and genomic sequence). Such nucleic acid molecules will consist of, consist essentially of, or comprise a nucleotide sequence that encodes one of the phosphatase peptides of the present invention, an allelic variant thereof, or an ortholog or paralog thereof.

As used herein, an "isolated" nucleic acid molecule is one that is separated from other nucleic acid present in the natural source of the nucleic acid. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. However, there can be some flanking nucleotide sequences, for example up to about 5KB, 4KB, 3KB, 2KB, or 1KB or less, particularly contiguous peptide encoding sequences and peptide encoding sequences within the same gene but separated by introns in the genomic sequence. The important point is that the nucleic acid is isolated from remote and unimportant flanking sequences such that it can be subjected to the specific manipulations described herein such as recombinant expression, preparation of probes and primers, and other uses specific to the nucleic acid sequences.

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Moreover, an "isolated" nucleic acid molecule, such as a transcript/cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or chemical precursors or other chemicals when chemically synthesized. However, the nucleic acid molecule can be fused to other coding or regulatory sequences and still be considered isolated.

For example, recombinant DNA molecules contained in a vector are considered isolated. Further examples of isolated DNA molecules include recombinant DNA molecules maintained in heterologous host cells or purified (partially or substantially) DNA molecules in solution. Isolated RNA molecules include *in vivo* or *in vitro* RNA transcripts of the isolated DNA molecules of the present invention. Isolated nucleic acid molecules according to the present invention further include such molecules produced synthetically.

Accordingly, the present invention provides nucleic acid molecules that consist of the nucleotide sequence shown in Figure 1 or 3 [SEQ ID NOS:1,4,6 (cDNA sequences) and SEQ ID NO:3 (genomic sequence)], or any nucleic acid molecule that encodes the protein provided in Figure 2, SEQ ID NOS:2,5,7. A nucleic acid molecule consists of a nucleotide sequence when the nucleotide sequence is the complete nucleotide sequence of the nucleic acid molecule.

The present invention further provides nucleic acid molecules that consist essentially of the nucleotide sequence shown in Figure 1 or 3 [SEQ ID NOS:1,4,6 (cDNA sequences) and SEQ ID NO:3 (genomic sequence)], or any nucleic acid molecule that encodes the protein provided in Figure 2, SEQ ID NOS:2,5,7. A nucleic acid molecule consists essentially of a nucleotide sequence when such a nucleotide sequence is present with only a few additional nucleic acid residues in the final nucleic acid molecule.

The present invention further provides nucleic acid molecules that comprise the nucleotide

sequences shown in Figure 1 or 3 [SEQ ID NOS:1,4,6 (cDNA sequences) and SEQ ID NO:3 (genomic sequence)], or any nucleic acid molecule that encodes the protein provided in Figure 2, SEQ ID NOS:2,5,7. A nucleic acid molecule comprises a nucleotide sequence when the nucleotide sequence is at least part of the final nucleotide sequence of the nucleic acid molecule. In such a fashion, the nucleic acid molecule can be only the nucleotide sequence or have additional nucleic acid residues, such as nucleic acid residues that are naturally associated with it or heterologous nucleotide sequences. Such a nucleic acid molecule can have a few additional nucleotides or can comprises several hundred or more additional nucleotides. A brief description of how various types of these nucleic acid molecules can be readily made/isolated is provided below.

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In Figures 1 and 3, both coding and non-coding sequences are provided. Because of the source of the present invention, humans genomic sequence (Figure 3) and cDNA/transcript sequences (Figure 1), the nucleic acid molecules in the Figures will contain genomic intronic sequences, 5' and 3' non-coding sequences, gene regulatory regions and non-coding intergenic sequences. In general such sequence features are either noted in Figures 1 and 3 or can readily be identified using computational tools known in the art. As discussed below, some of the non-coding regions, particularly gene regulatory elements such as promoters, are useful for a variety of purposes, e.g. control of heterologous gene expression, target for identifying gene activity modulating compounds, and are particularly claimed as fragments of the genomic sequence provided herein.

The isolated nucleic acid molecules can encode the mature protein plus additional amino or carboxyl-terminal amino acids, or amino acids interior to the mature peptide (when the mature form has more than one peptide chain, for instance). Such sequences may play a role in processing of a protein from precursor to a mature form, facilitate protein trafficking, prolong or shorten protein half-life or facilitate manipulation of a protein for assay or production, among other things. As generally is the case *in situ*, the additional amino acids may be processed away from the mature protein by cellular enzymes.

As mentioned above, the isolated nucleic acid molecules include, but are not limited to, the sequence encoding the phosphatase peptide alone, the sequence encoding the mature peptide and additional coding sequences, such as a leader or secretory sequence (e.g., a pre-pro or pro-protein sequence), the sequence encoding the mature peptide, with or without the additional coding sequences, plus additional non-coding sequences, for example introns and non-coding 5' and 3' sequences such as transcribed but non-translated sequences that play a role in transcription, mRNA processing (including splicing and polyadenylation signals), ribosome binding and stability of

mRNA. In addition, the nucleic acid molecule may be fused to a marker sequence encoding, for example, a peptide that facilitates purification.

Isolated nucleic acid molecules can be in the form of RNA, such as mRNA, or in the form DNA, including cDNA and genomic DNA obtained by cloning or produced by chemical synthetic techniques or by a combination thereof. The nucleic acid, especially DNA, can be double-stranded or single-stranded. Single-stranded nucleic acid can be the coding strand (sense strand) or the non-coding strand (anti-sense strand).

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The invention further provides nucleic acid molecules that encode fragments of the peptides of the present invention as well as nucleic acid molecules that encode obvious variants of the phosphatase proteins of the present invention that are described above. Such nucleic acid molecules may be naturally occurring, such as allelic variants (same locus), paralogs (different locus), and orthologs (different organism), or may be constructed by recombinant DNA methods or by chemical synthesis. Such non-naturally occurring variants may be made by mutagenesis techniques, including those applied to nucleic acid molecules, cells, or organisms. Accordingly, as discussed above, the variants can contain nucleotide substitutions, deletions, inversions and insertions. Variation can occur in either or both the coding and non-coding regions. The variations can produce both conservative and non-conservative amino acid substitutions.

The present invention further provides non-coding fragments of the nucleic acid molecules provided in Figures 1 and 3. Preferred non-coding fragments include, but are not limited to, promoter sequences, enhancer sequences, gene modulating sequences and gene termination sequences. Such fragments are useful in controlling heterologous gene expression and in developing screens to identify gene-modulating agents. A promoter can readily be identified as being 5' to the ATG start site in the genomic sequence provided in Figure 3.

A fragment comprises a contiguous nucleotide sequence greater than 12 or more nucleotides. Further, a fragment could at least 30, 40, 50, 100, 250 or 500 nucleotides in length. The length of the fragment will be based on its intended use. For example, the fragment can encode epitope bearing regions of the peptide, or can be useful as DNA probes and primers. Such fragments can be isolated using the known nucleotide sequence to synthesize an oligonucleotide probe. A labeled probe can then be used to screen a cDNA library, genomic DNA library, or mRNA to isolate nucleic acid corresponding to the coding region. Further, primers can be used in PCR reactions to clone specific regions of gene.

A probe/primer typically comprises substantially a purified oligonucleotide or oligonucleotide pair. The oligonucleotide typically comprises a region of nucleotide sequence that

hybridizes under stringent conditions to at least about 12, 20, 25, 40, 50 or more consecutive nucleotides.

Orthologs, homologs, and allelic variants can be identified using methods well known in the art. As described in the Peptide Section, these variants comprise a nucleotide sequence encoding a peptide that is typically 60-70%, 70-80%, 80-90%, and more typically at least about 90-95% or more homologous to the nucleotide sequence shown in the Figure sheets or a fragment of this sequence. Such nucleic acid molecules can readily be identified as being able to hybridize under moderate to stringent conditions, to the nucleotide sequence shown in the Figure sheets or a fragment of the sequence. Allelic variants can readily be determined by genetic locus of the encoding gene. The gene encoding the novel phosphatase proteins of the present invention is located on public BAC AP001885, which is known to be mapped to chromosome 11 (as indicated in Figure 3).

Figure 3 provides information on SNPs that have been found in the gene encoding the phosphatase proteins of the present invention. The following SNPs were identified: G577A, G1451A, and G2641A. G577A and G1451A are non-synonymous coding SNPs. Changes in the amino acid sequence caused by these SNPs is indicated in Figure 3 and can readily be determined using the universal genetic code and the protein sequence provided in Figure 2 as a reference. G2641A is 3' of the ORF and may affect control/regulatory elements.

As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences encoding a peptide at least 60-70% homologous to each other typically remain hybridized to each other. The conditions can be such that sequences at least about 60%, at least about 70%, or at least about 80% or more homologous to each other typically remain hybridized to each other. Such stringent conditions are known to those skilled in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. One example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50-65C. Examples of moderate to low stringency hybridization conditions are well known in the art.

30 Nucleic Acid Molecule Uses

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The nucleic acid molecules of the present invention are useful for probes, primers, chemical intermediates, and in biological assays. The nucleic acid molecules are useful as a hybridization

probe for messenger RNA, transcript/cDNA and genomic DNA to isolate full-length cDNA and genomic clones encoding the peptide described in Figure 2 and to isolate cDNA and genomic clones that correspond to variants (alleles, orthologs, etc.) producing the same or related peptides shown in Figure 2. As illustrated in Figure 3, the following SNP variations were identified: G577A, G1451A, and G2641A.

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The probe can correspond to any sequence along the entire length of the nucleic acid molecules provided in the Figures. Accordingly, it could be derived from 5' noncoding regions, the coding region, and 3' noncoding regions. However, as discussed, fragments are not to be construed as encompassing fragments disclosed prior to the present invention.

The nucleic acid molecules are also useful as primers for PCR to amplify any given region of a nucleic acid molecule and are useful to synthesize antisense molecules of desired length and sequence.

The nucleic acid molecules are also useful for constructing recombinant vectors. Such vectors include expression vectors that express a portion of, or all of, the peptide sequences. Vectors also include insertion vectors, used to integrate into another nucleic acid molecule sequence, such as into the cellular genome, to alter *in situ* expression of a gene and/or gene product. For example, an endogenous coding sequence can be replaced via homologous recombination with all or part of the coding region containing one or more specifically introduced mutations.

The nucleic acid molecules are also useful for expressing antigenic portions of the proteins.

The nucleic acid molecules are also useful as probes for determining the chromosomal positions of the nucleic acid molecules by means of *in situ* hybridization methods. The gene encoding the novel phosphatase proteins of the present invention is located on public BAC AP001885, which is known to be mapped to chromosome 11 (as indicated in Figure 3).

The nucleic acid molecules are also useful in making vectors containing the gene regulatory regions of the nucleic acid molecules of the present invention.

The nucleic acid molecules are also useful for designing ribozymes corresponding to all, or a part, of the mRNA produced from the nucleic acid molecules described herein.

The nucleic acid molecules are also useful for making vectors that express part, or all, of the peptides.

The nucleic acid molecules are also useful for constructing host cells expressing a part, or all, of the nucleic acid molecules and peptides.

The nucleic acid molecules are also useful for constructing transgenic animals expressing all, or a part, of the nucleic acid molecules and peptides.

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The nucleic acid molecules are also useful as hybridization probes for determining the presence, level, form and distribution of nucleic acid expression. Experimental data as provided in Figure 1 indicates that splice form 1 of the phosphatase protein of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Specifically, a virtual northern blot shows expression in pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, and ovary tumor tissue. In addition, PCR-based tissue screening panels indicate expression in the brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Accordingly, the probes can be used to detect the presence of, or to determine levels of, a specific nucleic acid molecule in cells, tissues, and in organisms. The nucleic acid whose level is determined can be DNA or RNA. Accordingly, probes corresponding to the peptides described herein can be used to assess expression and/or gene copy number in a given cell, tissue, or organism. These uses are relevant for diagnosis of disorders involving an increase or decrease in phosphatase protein expression relative to normal results.

In vitro techniques for detection of mRNA include Northern hybridizations and in situ hybridizations. In vitro techniques for detecting DNA includes Southern hybridizations and in situ hybridization.

Probes can be used as a part of a diagnostic test kit for identifying cells or tissues that express a phosphatase protein, such as by measuring a level of a phosphatase-encoding nucleic acid in a sample of cells from a subject e.g., mRNA or genomic DNA, or determining if a phosphatase gene has been mutated. Experimental data as provided in Figure 1 indicates that splice form 1 of the phosphatase protein of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Specifically, a virtual northern blot shows expression in pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, and ovary tumor tissue. In addition, PCR-based tissue screening panels indicate expression in the brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid.

Nucleic acid expression assays are useful for drug screening to identify compounds that

modulate phosphatase nucleic acid expression.

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The invention thus provides a method for identifying a compound that can be used to treat a disorder associated with nucleic acid expression of the phosphatase gene, particularly biological and pathological processes that are mediated by the phosphatase in cells and tissues that express it. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. The method typically includes assaying the ability of the compound to modulate the expression of the phosphatase nucleic acid and thus identifying a compound that can be used to treat a disorder characterized by undesired phosphatase nucleic acid expression. The assays can be performed in cell-based and cell-free systems. Cell-based assays include cells naturally expressing the phosphatase nucleic acid or recombinant cells genetically engineered to express specific nucleic acid sequences.

The assay for phosphatase nucleic acid expression can involve direct assay of nucleic acid levels, such as mRNA levels, or on collateral compounds involved in the signal pathway. Further, the expression of genes that are up- or down-regulated in response to the phosphatase protein signal pathway can also be assayed. In this embodiment the regulatory regions of these genes can be operably linked to a reporter gene such as luciferase.

Thus, modulators of phosphatase gene expression can be identified in a method wherein a cell is contacted with a candidate compound and the expression of mRNA determined. The level of expression of phosphatase mRNA in the presence of the candidate compound is compared to the level of expression of phosphatase mRNA in the absence of the candidate compound. The candidate compound can then be identified as a modulator of nucleic acid expression based on this comparison and be used, for example to treat a disorder characterized by aberrant nucleic acid expression. When expression of mRNA is statistically significantly greater in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of nucleic acid expression. When nucleic acid expression is statistically significantly less in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of nucleic acid expression.

The invention further provides methods of treatment, with the nucleic acid as a target, using a compound identified through drug screening as a gene modulator to modulate phosphatase nucleic acid expression in cells and tissues that express the phosphatase. Experimental data as provided in

Figure 1 indicates that splice form 1 of the phosphatase protein of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Specifically, a virtual northern blot shows expression in pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, and ovary tumor tissue. In addition, PCR-based tissue screening panels indicate expression in the brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Modulation includes both upregulation (i.e. activation or agonization) or down-regulation (suppression or antagonization) or nucleic acid expression.

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Alternatively, a modulator for phosphatase nucleic acid expression can be a small molecule or drug identified using the screening assays described herein as long as the drug or small molecule inhibits the phosphatase nucleic acid expression in the cells and tissues that express the protein. Experimental data as provided in Figure 1 indicates that splice form 1 of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain.

The nucleic acid molecules are also useful for monitoring the effectiveness of modulating compounds on the expression or activity of the phosphatase gene in clinical trials or in a treatment regimen. Thus, the gene expression pattern can serve as a barometer for the continuing effectiveness of treatment with the compound, particularly with compounds to which a patient can develop resistance. The gene expression pattern can also serve as a marker indicative of a physiological response of the affected cells to the compound. Accordingly, such monitoring would allow either increased administration of the compound or the administration of alternative compounds to which the patient has not become resistant. Similarly, if the level of nucleic acid expression falls below a desirable level, administration of the compound could be commensurately decreased.

The nucleic acid molecules are also useful in diagnostic assays for qualitative changes in phosphatase nucleic acid expression, and particularly in qualitative changes that lead to pathology. The nucleic acid molecules can be used to detect mutations in phosphatase genes and gene expression products such as mRNA. The nucleic acid molecules can be used as hybridization

probes to detect naturally occurring genetic mutations in the phosphatase gene and thereby to determine whether a subject with the mutation is at risk for a disorder caused by the mutation. Mutations include deletion, addition, or substitution of one or more nucleotides in the gene, chromosomal rearrangement, such as inversion or transposition, modification of genomic DNA, such as aberrant methylation patterns or changes in gene copy number, such as amplification. Detection of a mutated form of the phosphatase gene associated with a dysfunction provides a diagnostic tool for an active disease or susceptibility to disease when the disease results from overexpression, underexpression, or altered expression of a phosphatase protein.

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Individuals carrying mutations in the phosphatase gene can be detected at the nucleic acid level by a variety of techniques. Figure 3 provides information on SNPs that have been found in the gene encoding the phosphatase proteins of the present invention. The following SNPs were identified: G577A, G1451A, and G2641A. G577A and G1451A are non-synonymous coding SNPs. Changes in the amino acid sequence caused by these SNPs is indicated in Figure 3 and can readily be determined using the universal genetic code and the protein sequence provided in Figure 2 as a reference. G2641A is 3' of the ORF and may affect control/regulatory elements. The gene encoding the novel phosphatase proteins of the present invention is located on public BAC AP001885, which is known to be mapped to chromosome 11 (as indicated in Figure 3). Genomic DNA can be analyzed directly or can be amplified by using PCR prior to analysis. RNA or cDNA can be used in the same way. In some uses, detection of the mutation involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, e.g. U.S. Patent Nos. 4,683,195 and 4,683,202), such as anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, e.g., Landegran et al., Science 241:1077-1080 (1988); and Nakazawa et al., PNAS 91:360-364 (1994)), the latter of which can be particularly useful for detecting point mutations in the gene (see Abravaya et al., Nucleic Acids Res. 23:675-682 (1995)). This method can include the steps of collecting a sample of cells from a patient, isolating nucleic acid (e.g., genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or more primers which specifically hybridize to a gene under conditions such that hybridization and amplification of the gene (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. Deletions and insertions can be detected by a change in size of the amplified product compared to the normal genotype. Point mutations can be identified by hybridizing amplified DNA to normal RNA or antisense DNA sequences.

Alternatively, mutations in a phosphatase gene can be directly identified, for example, by

alterations in restriction enzyme digestion patterns determined by gel electrophoresis.

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Further, sequence-specific ribozymes (U.S. Patent No. 5,498,531) can be used to score for the presence of specific mutations by development or loss of a ribozyme cleavage site. Perfectly matched sequences can be distinguished from mismatched sequences by nuclease cleavage digestion assays or by differences in melting temperature.

Sequence changes at specific locations can also be assessed by nuclease protection assays such as RNase and S1 protection or the chemical cleavage method. Furthermore, sequence differences between a mutant phosphatase gene and a wild-type gene can be determined by direct DNA sequencing. A variety of automated sequencing procedures can be utilized when performing the diagnostic assays (Naeve, C.W., (1995) *Biotechniques 19*:448), including sequencing by mass spectrometry (see, e.g., PCT International Publication No. WO 94/16101; Cohen et al., Adv. Chromatogr. 36:127-162 (1996); and Griffin et al., Appl. Biochem. Biotechnol. 38:147-159 (1993)).

Other methods for detecting mutations in the gene include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or RNA/DNA duplexes (Myers et al., Science 230:1242 (1985)); Cotton et al., PNAS 85:4397 (1988); Saleeba et al., Meth. Enzymol. 217:286-295 (1992)), electrophoretic mobility of mutant and wild type nucleic acid is compared (Orita et al., PNAS 86:2766 (1989); Cotton et al., Mutat. Res. 285:125-144 (1993); and Hayashi et al., Genet. Anal. Tech. Appl. 9:73-79 (1992)), and movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (Myers et al., Nature 313:495 (1985)). Examples of other techniques for detecting point mutations include selective oligonucleotide hybridization, selective amplification, and selective primer extension.

The nucleic acid molecules are also useful for testing an individual for a genotype that while not necessarily causing the disease, nevertheless affects the treatment modality. Thus, the nucleic acid molecules can be used to study the relationship between an individual's genotype and the individual's response to a compound used for treatment (pharmacogenomic relationship). Accordingly, the nucleic acid molecules described herein can be used to assess the mutation content of the phosphatase gene in an individual in order to select an appropriate compound or dosage regimen for treatment. Figure 3 provides information on SNPs that have been found in the gene encoding the phosphatase proteins of the present invention. The following SNPs were identified: G577A, G1451A, and G2641A. G577A and G1451A are non-synonymous coding SNPs. Changes in the amino acid sequence caused by these SNPs is indicated in Figure 3 and can readily be determined using the universal genetic code and the protein sequence provided in Figure 2 as a

reference. G2641A is 3' of the ORF and may affect control/regulatory elements.

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Thus nucleic acid molecules displaying genetic variations that affect treatment provide a diagnostic target that can be used to tailor treatment in an individual. Accordingly, the production of recombinant cells and animals containing these polymorphisms allow effective clinical design of treatment compounds and dosage regimens.

The nucleic acid molecules are thus useful as antisense constructs to control phosphatase gene expression in cells, tissues, and organisms. A DNA antisense nucleic acid molecule is designed to be complementary to a region of the gene involved in transcription, preventing transcription and hence production of phosphatase protein. An antisense RNA or DNA nucleic acid molecule would hybridize to the mRNA and thus block translation of mRNA into phosphatase protein.

Alternatively, a class of antisense molecules can be used to inactivate mRNA in order to decrease expression of phosphatase nucleic acid. Accordingly, these molecules can treat a disorder characterized by abnormal or undesired phosphatase nucleic acid expression. This technique involves cleavage by means of ribozymes containing nucleotide sequences complementary to one or more regions in the mRNA that attenuate the ability of the mRNA to be translated. Possible regions include coding regions and particularly coding regions corresponding to the catalytic and other functional activities of the phosphatase protein, such as substrate binding.

The nucleic acid molecules also provide vectors for gene therapy in patients containing cells that are aberrant in phosphatase gene expression. Thus, recombinant cells, which include the patient's cells that have been engineered *ex vivo* and returned to the patient, are introduced into an individual where the cells produce the desired phosphatase protein to treat the individual.

The invention also encompasses kits for detecting the presence of a phosphatase nucleic acid in a biological sample. Experimental data as provided in Figure 1 indicates that splice form 1 of the phosphatase protein of the present invention is expressed in humans in the pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, ovary tumor tissue, brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. Additionally, splice form 3 is expressed in fetal brain. Specifically, a virtual northern blot shows expression in pancreas, colon and pancreas adenocarcinomas, pancreas epitheliod carcinomas, lung large cell carcinomas, renal cell carcinomas, placenta choriocarcinomas, and ovary tumor tissue. In addition, PCR-based tissue screening panels indicate expression in the brain (including fetal), heart (including fetal), kidney (including fetal), uterus, and thyroid. For example, the kit can comprise reagents such as a labeled

or labelable nucleic acid or agent capable of detecting phosphatase nucleic acid in a biological sample; means for determining the amount of phosphatase nucleic acid in the sample; and means for comparing the amount of phosphatase nucleic acid in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect phosphatase protein mRNA or DNA.

Nucleic Acid Arrays

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The present invention further provides nucleic acid detection kits, such as arrays or microarrays of nucleic acid molecules that are based on the sequence information provided in Figures 1 and 3 (SEQ ID NOS:1,3,4,6).

As used herein "Arrays" or "Microarrays" refers to an array of distinct polynucleotides or oligonucleotides synthesized on a substrate, such as paper, nylon or other type of membrane, filter, chip, glass slide, or any other suitable solid support. In one embodiment, the microarray is prepared and used according to the methods described in US Patent 5,837,832, Chee *et al.*, PCT application W095/11995 (Chee *et al.*), Lockhart, D. J. *et al.* (1996; Nat. Biotech. 14: 1675-1680) and Schena, M. *et al.* (1996; Proc. Natl. Acad. Sci. 93: 10614-10619), all of which are incorporated herein in their entirety by reference. In other embodiments, such arrays are produced by the methods described by Brown *et al.*, US Patent No. 5,807,522.

The microarray or detection kit is preferably composed of a large number of unique, single-stranded nucleic acid sequences, usually either synthetic antisense oligonucleotides or fragments of cDNAs, fixed to a solid support. The oligonucleotides are preferably about 6-60 nucleotides in length, more preferably 15-30 nucleotides in length, and most preferably about 20-25 nucleotides in length. For a certain type of microarray or detection kit, it may be preferable to use oligonucleotides that are only 7-20 nucleotides in length. The microarray or detection kit may contain oligonucleotides that cover the known 5', or 3', sequence, sequential oligonucleotides which cover the full length sequence; or unique oligonucleotides selected from particular areas along the length of the sequence. Polynucleotides used in the microarray or detection kit may be oligonucleotides that are specific to a gene or genes of interest.

In order to produce oligonucleotides to a known sequence for a microarray or detection kit, the gene(s) of interest (or an ORF identified from the contigs of the present invention) is typically examined using a computer algorithm which starts at the 5' or at the 3' end of the nucleotide sequence. Typical algorithms will then identify oligomers of defined length that are

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unique to the gene, have a GC content within a range suitable for hybridization, and lack predicted secondary structure that may interfere with hybridization. In certain situations it may be appropriate to use pairs of oligonucleotides on a microarray or detection kit. The "pairs" will be identical, except for one nucleotide that preferably is located in the center of the sequence. The second oligonucleotide in the pair (mismatched by one) serves as a control. The number of oligonucleotide pairs may range from two to one million. The oligomers are synthesized at designated areas on a substrate using a light-directed chemical process. The substrate may be paper, nylon or other type of membrane, filter, chip, glass slide or any other suitable solid support.

In another aspect, an oligonucleotide may be synthesized on the surface of the substrate by using a chemical coupling procedure and an ink jet application apparatus, as described in PCT application W095/251116 (Baldeschweiler *et al.*) which is incorporated herein in its entirety by reference. In another aspect, a "gridded" array analogous to a dot (or slot) blot may be used to arrange and link cDNA fragments or oligonucleotides to the surface of a substrate using a vacuum system, thermal, UV, mechanical or chemical bonding procedures. An array, such as those described above, may be produced by hand or by using available devices (slot blot or dot blot apparatus), materials (any suitable solid support), and machines (including robotic instruments), and may contain 8, 24, 96, 384, 1536, 6144 or more oligonucleotides, or any other number between two and one million which lends itself to the efficient use of commercially available instrumentation.

In order to conduct sample analysis using a microarray or detection kit, the RNA or DNA from a biological sample is made into hybridization probes. The mRNA is isolated, and cDNA is produced and used as a template to make antisense RNA (aRNA). The aRNA is amplified in the presence of fluorescent nucleotides, and labeled probes are incubated with the microarray or detection kit so that the probe sequences hybridize to complementary oligonucleotides of the microarray or detection kit. Incubation conditions are adjusted so that hybridization occurs with precise complementary matches or with various degrees of less complementarity. After removal of nonhybridized probes, a scanner is used to determine the levels and patterns of fluorescence. The scanned images are examined to determine degree of complementarity and the relative abundance of each oligonucleotide sequence on the microarray or detection kit. The biological samples may be obtained from any bodily fluids (such as blood, urine, saliva, phlegm, gastric juices, etc.), cultured cells, biopsies, or other tissue preparations. A detection system may be used to measure the absence, presence, and amount of hybridization for all of the distinct

sequences simultaneously. This data may be used for large-scale correlation studies on the sequences, expression patterns, mutations, variants, or polymorphisms among samples.

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Using such arrays, the present invention provides methods to identify the expression of the phosphatase proteins/peptides of the present invention. In detail, such methods comprise incubating a test sample with one or more nucleic acid molecules and assaying for binding of the nucleic acid molecule with components within the test sample. Such assays will typically involve arrays comprising many genes, at least one of which is a gene of the present invention and or alleles of the phosphatase gene of the present invention. Figure 3 provides information on SNPs that have been found in the gene encoding the phosphatase proteins of the present invention. The following SNPs were identified: G577A, G1451A, and G2641A. G577A and G1451A are non-synonymous coding SNPs. Changes in the amino acid sequence caused by these SNPs is indicated in Figure 3 and can readily be determined using the universal genetic code and the protein sequence provided in Figure 2 as a reference. G2641A is 3° of the ORF and may affect control/regulatory elements.

Conditions for incubating a nucleic acid molecule with a test sample vary. Incubation conditions depend on the format employed in the assay, the detection methods employed, and the type and nature of the nucleic acid molecule used in the assay. One skilled in the art will recognize that any one of the commonly available hybridization, amplification or array assay formats can readily be adapted to employ the novel fragments of the Human genome disclosed herein. Examples of such assays can be found in Chard, T, An Introduction to Radioimmunoassay and Related Techniques, Elsevier Science Publishers, Amsterdam, The Netherlands (1986); Bullock, G. R. et al., Techniques in Immunocytochemistry, Academic Press, Orlando, FL Vol. 1 (1 982), Vol. 2 (1983), Vol. 3 (1985); Tijssen, P., Practice and Theory of Enzyme Immunoassays: Laboratory Techniques in Biochemistry and Molecular Biology, Elsevier Science Publishers, Amsterdam, The Netherlands (1985).

The test samples of the present invention include cells, protein or membrane extracts of cells. The test sample used in the above-described method will vary based on the assay format, nature of the detection method and the tissues, cells or extracts used as the sample to be assayed. Methods for preparing nucleic acid extracts or of cells are well known in the art and can be readily be adapted in order to obtain a sample that is compatible with the system utilized.

In another embodiment of the present invention, kits are provided which contain the necessary reagents to carry out the assays of the present invention.

Specifically, the invention provides a compartmentalized kit to receive, in close

confinement, one or more containers which comprises: (a) a first container comprising one of the nucleic acid molecules that can bind to a fragment of the Human genome disclosed herein; and (b) one or more other containers comprising one or more of the following: wash reagents, reagents capable of detecting presence of a bound nucleic acid.

In detail, a compartmentalized kit includes any kit in which reagents are contained in separate containers. Such containers include small glass containers, plastic containers, strips of plastic, glass or paper, or arraying material such as silica. Such containers allows one to efficiently transfer reagents from one compartment to another compartment such that the samples and reagents are not cross-contaminated, and the agents or solutions of each container can be added in a quantitative fashion from one compartment to another. Such containers will include a container which will accept the test sample, a container which contains the nucleic acid probe, containers which contain wash reagents (such as phosphate buffered saline, Tris-buffers, etc.), and containers which contain the reagents used to detect the bound probe. One skilled in the art will readily recognize that the previously unidentified phosphatase gene of the present invention can be routinely identified using the sequence information disclosed herein can be readily incorporated into one of the established kit formats which are well known in the art, particularly expression arrays.

Vectors/host cells

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The invention also provides vectors containing the nucleic acid molecules described herein. The term "vector" refers to a vehicle, preferably a nucleic acid molecule, which can transport the nucleic acid molecules. When the vector is a nucleic acid molecule, the nucleic acid molecules are covalently linked to the vector nucleic acid. With this aspect of the invention, the vector includes a plasmid, single or double stranded phage, a single or double stranded RNA or DNA viral vector, or artificial chromosome, such as a BAC, PAC, YAC, OR MAC.

A vector can be maintained in the host cell as an extrachromosomal element where it replicates and produces additional copies of the nucleic acid molecules. Alternatively, the vector may integrate into the host cell genome and produce additional copies of the nucleic acid molecules when the host cell replicates.

The invention provides vectors for the maintenance (cloning vectors) or vectors for expression (expression vectors) of the nucleic acid molecules. The vectors can function in prokaryotic or eukaryotic cells or in both (shuttle vectors).

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Expression vectors contain cis-acting regulatory regions that are operably linked in the vector to the nucleic acid molecules such that transcription of the nucleic acid molecules is allowed in a host cell. The nucleic acid molecules can be introduced into the host cell with a separate nucleic acid molecule capable of affecting transcription. Thus, the second nucleic acid molecule may provide a trans-acting factor interacting with the cis-regulatory control region to allow transcription of the nucleic acid molecules from the vector. Alternatively, a trans-acting factor may be supplied by the host cell. Finally, a trans-acting factor can be produced from the vector itself. It is understood, however, that in some embodiments, transcription and/or translation of the nucleic acid molecules can occur in a cell-free system.

The regulatory sequence to which the nucleic acid molecules described herein can be operably linked include promoters for directing mRNA transcription. These include, but are not limited to, the left promoter from bacteriophage λ , the lac, TRP, and TAC promoters from *E. coli*, the early and late promoters from SV40, the CMV immediate early promoter, the adenovirus early and late promoters, and retrovirus long-terminal repeats.

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In addition to control regions that promote transcription, expression vectors may also include regions that modulate transcription, such as repressor binding sites and enhancers. Examples include the SV40 enhancer, the cytomegalovirus immediate early enhancer, polyoma enhancer, adenovirus enhancers, and retrovirus LTR enhancers.

In addition to containing sites for transcription initiation and control, expression vectors can also contain sequences necessary for transcription termination and, in the transcribed region a ribosome binding site for translation. Other regulatory control elements for expression include initiation and termination codons as well as polyadenylation signals. The person of ordinary skill in the art would be aware of the numerous regulatory sequences that are useful in expression vectors. Such regulatory sequences are described, for example, in Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual. 2nd. ed.*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, (1989).

A variety of expression vectors can be used to express a nucleic acid molecule. Such vectors include chromosomal, episomal, and virus-derived vectors, for example vectors derived from bacterial plasmids, from bacteriophage, from yeast episomes, from yeast chromosomal elements, including yeast artificial chromosomes, from viruses such as baculoviruses, papovaviruses such as SV40, Vaccinia viruses, adenoviruses, poxviruses, pseudorabies viruses, and retroviruses. Vectors may also be derived from combinations of these sources such as those derived from plasmid and bacteriophage genetic elements, e.g. cosmids and phagemids. Appropriate

cloning and expression vectors for prokaryotic and eukaryotic hosts are described in Sambrook et al., Molecular Cloning: A Laboratory Manual. 2nd. ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, (1989).

The regulatory sequence may provide constitutive expression in one or more host cells (i.e. tissue specific) or may provide for inducible expression in one or more cell types such as by temperature, nutrient additive, or exogenous factor such as a hormone or other ligand. A variety of vectors providing for constitutive and inducible expression in prokaryotic and eukaryotic hosts are well known to those of ordinary skill in the art.

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The nucleic acid molecules can be inserted into the vector nucleic acid by well-known methodology. Generally, the DNA sequence that will ultimately be expressed is joined to an expression vector by cleaving the DNA sequence and the expression vector with one or more restriction enzymes and then ligating the fragments together. Procedures for restriction enzyme digestion and ligation are well known to those of ordinary skill in the art.

The vector containing the appropriate nucleic acid molecule can be introduced into an appropriate host cell for propagation or expression using well-known techniques. Bacterial cells include, but are not limited to, *E. coli*, *Streptomyces*, and *Salmonella typhimurium*. Eukaryotic cells include, but are not limited to, yeast, insect cells such as *Drosophila*, animal cells such as COS and CHO cells, and plant cells.

As described herein, it may be desirable to express the peptide as a fusion protein. Accordingly, the invention provides fusion vectors that allow for the production of the peptides. Fusion vectors can increase the expression of a recombinant protein, increase the solubility of the recombinant protein, and aid in the purification of the protein by acting for example as a ligand for affinity purification. A proteolytic cleavage site may be introduced at the junction of the fusion moiety so that the desired peptide can ultimately be separated from the fusion moiety. Proteolytic enzymes include, but are not limited to, factor Xa, thrombin, and enterophosphatase. Typical fusion expression vectors include pGEX (Smith et al., Gene 67:31-40 (1988)), pMAL (New England Biolabs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ) which fuse glutathione Stransferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein. Examples of suitable inducible non-fusion E. coli expression vectors include pTrc (Amann et al., Gene 69:301-315 (1988)) and pET 11d (Studier et al., Gene Expression Technology: Methods in Enzymology 185:60-89 (1990)).

Recombinant protein expression can be maximized in host bacteria by providing a genetic background wherein the host cell has an impaired capacity to proteolytically cleave the recombinant

protein. (Gottesman, S., Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, California (1990) 119-128). Alternatively, the sequence of the nucleic acid molecule of interest can be altered to provide preferential codon usage for a specific host cell, for example E. coli. (Wada et al., Nucleic Acids Res. 20:2111-2118 (1992)).

The nucleic acid molecules can also be expressed by expression vectors that are operative in yeast. Examples of vectors for expression in yeast e.g., *S. cerevisiae* include pYepSec1 (Baldari, *et al.*, *EMBO J. 6*:229-234 (1987)), pMFa (Kurjan *et al.*, *Cell 30*:933-943(1982)), pJRY88 (Schultz *et al.*, *Gene 54*:113-123 (1987)), and pYES2 (Invitrogen Corporation, San Diego, CA).

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The nucleic acid molecules can also be expressed in insect cells using, for example, baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (e.g., Sf 9 cells) include the pAc series (Smith *et al.*, *Mol. Cell Biol. 3*:2156-2165 (1983)) and the pVL series (Lucklow *et al.*, *Virology 170*:31-39 (1989)).

In certain embodiments of the invention, the nucleic acid molecules described herein are expressed in mammalian cells using mammalian expression vectors. Examples of mammalian expression vectors include pCDM8 (Seed, B. *Nature 329*:840(1987)) and pMT2PC (Kaufman *et al.*, *EMBO J. 6*:187-195 (1987)).

The expression vectors listed herein are provided by way of example only of the well-known vectors available to those of ordinary skill in the art that would be useful to express the nucleic acid molecules. The person of ordinary skill in the art would be aware of other vectors suitable for maintenance propagation or expression of the nucleic acid molecules described herein. These are found for example in Sambrook, J., Fritsh, E. F., and Maniatis, T. *Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory,* Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

The invention also encompasses vectors in which the nucleic acid sequences described herein are cloned into the vector in reverse orientation, but operably linked to a regulatory sequence that permits transcription of antisense RNA. Thus, an antisense transcript can be produced to all, or to a portion, of the nucleic acid molecule sequences described herein, including both coding and non-coding regions. Expression of this antisense RNA is subject to each of the parameters described above in relation to expression of the sense RNA (regulatory sequences, constitutive or inducible expression, tissue-specific expression).

The invention also relates to recombinant host cells containing the vectors described herein. Host cells therefore include prokaryotic cells, lower eukaryotic cells such as yeast, other eukaryotic cells such as insect cells, and higher eukaryotic cells such as mammalian cells.

The recombinant host cells are prepared by introducing the vector constructs described herein into the cells by techniques readily available to the person of ordinary skill in the art. These include, but are not limited to, calcium phosphate transfection, DEAE-dextran-mediated transfection, cationic lipid-mediated transfection, electroporation, transduction, infection, lipofection, and other techniques such as those found in Sambrook, et al. (Molecular Cloning: A Laboratory Manual. 2nd, ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989).

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Host cells can contain more than one vector. Thus, different nucleotide sequences can be introduced on different vectors of the same cell. Similarly, the nucleic acid molecules can be introduced either alone or with other nucleic acid molecules that are not related to the nucleic acid molecules such as those providing trans-acting factors for expression vectors. When more than one vector is introduced into a cell, the vectors can be introduced independently, co-introduced or joined to the nucleic acid molecule vector.

In the case of bacteriophage and viral vectors, these can be introduced into cells as packaged or encapsulated virus by standard procedures for infection and transduction. Viral vectors can be replication-competent or replication-defective. In the case in which viral replication is defective, replication will occur in host cells providing functions that complement the defects.

Vectors generally include selectable markers that enable the selection of the subpopulation of cells that contain the recombinant vector constructs. The marker can be contained in the same vector that contains the nucleic acid molecules described herein or may be on a separate vector. Markers include tetracycline or ampicillin-resistance genes for prokaryotic host cells and dihydrofolate reductase or neomycin resistance for eukaryotic host cells. However, any marker that provides selection for a phenotypic trait will be effective.

While the mature proteins can be produced in bacteria, yeast, mammalian cells, and other cells under the control of the appropriate regulatory sequences, cell- free transcription and translation systems can also be used to produce these proteins using RNA derived from the DNA constructs described herein.

Where secretion of the peptide is desired, which is difficult to achieve with multitransmembrane domain containing proteins such as phosphatases, appropriate secretion signals are incorporated into the vector. The signal sequence can be endogenous to the peptides or heterologous to these peptides.

Where the peptide is not secreted into the medium, which is typically the case with phosphatases, the protein can be isolated from the host cell by standard disruption procedures,

including freeze thaw, sonication, mechanical disruption, use of lysing agents and the like. The peptide can then be recovered and purified by well-known purification methods including ammonium sulfate precipitation, acid extraction, anion or cationic exchange chromatography, phosphocellulose chromatography, hydrophobic-interaction chromatography, affinity chromatography, hydroxylapatite chromatography, lectin chromatography, or high performance liquid chromatography.

It is also understood that depending upon the host cell in recombinant production of the peptides described herein, the peptides can have various glycosylation patterns, depending upon the cell, or maybe non-glycosylated as when produced in bacteria. In addition, the peptides may include an initial modified methionine in some cases as a result of a host-mediated process.

Uses of vectors and host cells

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The recombinant host cells expressing the peptides described herein have a variety of uses. First, the cells are useful for producing a phosphatase protein or peptide that can be further purified to produce desired amounts of phosphatase protein or fragments. Thus, host cells containing expression vectors are useful for peptide production.

Host cells are also useful for conducting cell-based assays involving the phosphatase protein or phosphatase protein fragments, such as those described above as well as other formats known in the art. Thus, a recombinant host cell expressing a native phosphatase protein is useful for assaying compounds that stimulate or inhibit phosphatase protein function.

Host cells are also useful for identifying phosphatase protein mutants in which these functions are affected. If the mutants naturally occur and give rise to a pathology, host cells containing the mutations are useful to assay compounds that have a desired effect on the mutant phosphatase protein (for example, stimulating or inhibiting function) which may not be indicated by their effect on the native phosphatase protein.

Genetically engineered host cells can be further used to produce non-human transgenic animals. A transgenic animal is preferably a mammal, for example a rodent, such as a rat or mouse, in which one or more of the cells of the animal include a transgene. A transgene is exogenous DNA which is integrated into the genome of a cell from which a transgenic animal develops and which remains in the genome of the mature animal in one or more cell types or tissues of the transgenic animal. These animals are useful for studying the function of a phosphatase protein and identifying and evaluating modulators of phosphatase protein activity. Other examples of transgenic animals

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include non-human primates, sheep, dogs, cows, goats, chickens, and amphibians.

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A transgenic animal can be produced by introducing nucleic acid into the male pronuclei of a fertilized oocyte, e.g., by microinjection, retroviral infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. Any of the phosphatase protein nucleotide sequences can be introduced as a transgene into the genome of a non-human animal, such as a mouse.

Any of the regulatory or other sequences useful in expression vectors can form part of the transgenic sequence. This includes intronic sequences and polyadenylation signals, if not already included. A tissue-specific regulatory sequence(s) can be operably linked to the transgene to direct expression of the phosphatase protein to particular cells.

Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are described, for example, in U.S. Patent Nos. 4,736,866 and 4,870,009, both by Leder et al., U.S. Patent No. 4,873,191 by Wagner et al. and in Hogan, B., Manipulating the Mouse Embryo, (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986). Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified based upon the presence of the transgene in its genome and/or expression of transgenic mRNA in tissues or cells of the animals. A transgenic founder animal can then be used to breed additional animals carrying the transgene. Moreover, transgenic animals carrying a transgene can further be bred to other transgenic animals carrying other transgenes. A transgenic animal also includes animals in which the entire animal or tissues in the animal have been produced using the homologously recombinant host cells described herein.

In another embodiment, transgenic non-human animals can be produced which contain selected systems that allow for regulated expression of the transgene. One example of such a system is the *cre/loxP* recombinase system of bacteriophage P1. For a description of the *cre/loxP* recombinase system, see, e.g., Lakso *et al. PNAS 89*:6232-6236 (1992). Another example of a recombinase system is the FLP recombinase system of *S. cerevisiae* (O'Gorman *et al. Science 251*:1351-1355 (1991). If a *cre/loxP* recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the *Cre* recombinase and a selected protein is required. Such animals can be provided through the construction of "double" transgenic animals, e.g., by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut, I. et al. Nature 385:810-813 (1997) and PCT

International Publication Nos. WO 97/07668 and WO 97/07669. In brief, a cell, e.g., a somatic cell, from the transgenic animal can be isolated and induced to exit the growth cycle and enter G₀ phase. The quiescent cell can then be fused, e.g., through the use of electrical pulses, to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The reconstructed oocyte is then cultured such that it develops to morula or blastocyst and then transferred to pseudopregnant female foster animal. The offspring born of this female foster animal will be a clone of the animal from which the cell, e.g., the somatic cell, is isolated.

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Transgenic animals containing recombinant cells that express the peptides described herein are useful to conduct the assays described herein in an *in vivo* context. Accordingly, the various physiological factors that are present *in vivo* and that could effect substrate binding, kinase protein activation, and signal transduction, may not be evident from *in vitro* cell-free or cell-based assays. Accordingly, it is useful to provide non-human transgenic animals to assay *in vivo* phosphatase protein function, including substrate interaction, the effect of specific mutant phosphatase proteins on phosphatase protein function and substrate interaction, and the effect of chimeric phosphatase proteins. It is also possible to assess the effect of null mutations, that is mutations that substantially or completely eliminate one or more phosphatase protein functions.

All publications and patents mentioned in the above specification are herein incorporated by reference. Various modifications and variations of the described method and system of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the above-described modes for carrying out the invention which are obvious to those skilled in the field of molecular biology or related fields are intended to be within the scope of the following claims.

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Claims

That which is claimed is:

1. An isolated peptide consisting of an amino acid sequence selected from the group consisting of:

- (a) an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7;
- (b) an amino acid sequence of an allelic variant of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said allelic variant is encoded by a nucleic acid molecule that hybridizes under stringent conditions to the opposite strand of a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6;
- (c) an amino acid sequence of an ortholog of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said ortholog is encoded by a nucleic acid molecule that hybridizes under stringent conditions to the opposite strand of a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6; and
- (d) a fragment of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said fragment comprises at least 10 contiguous amino acids.
- 2. An isolated peptide comprising an amino acid sequence selected from the group consisting of:
- (a) an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7;
- (b) an amino acid sequence of an allelic variant of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said allelic variant is encoded by a nucleic acid molecule that hybridizes under stringent conditions to the opposite strand of a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6;
 - (c) an amino acid sequence of an ortholog of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said ortholog is encoded by a nucleic acid molecule that hybridizes under stringent conditions to the opposite strand of a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6; and
 - (d) a fragment of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said fragment comprises at least 10 contiguous amino acids.

- 3. An isolated antibody that selectively binds to a peptide of claim 2.
- 4. An isolated nucleic acid molecule consisting of a nucleotide sequence selected from the group consisting of:
- (a) a nucleotide sequence that encodes an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7;
- (b) a nucleotide sequence that encodes of an allelic variant of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said nucleotide sequence hybridizes under stringent conditions to the opposite strand of a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6;
- (c) a nucleotide sequence that encodes an ortholog of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said nucleotide sequence hybridizes under stringent conditions to the opposite strand of a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6;
- (d) a nucleotide sequence that encodes a fragment of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said fragment comprises at least 10 contiguous amino acids; and
- (e) a nucleotide sequence that is the complement of a nucleotide sequence of (a)-(d).
- 5. An isolated nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of:
- (a) a nucleotide sequence that encodes an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7;
- (b) a nucleotide sequence that encodes of an allelic variant of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said nucleotide sequence hybridizes under stringent conditions to the opposite strand of a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6;
- (c) a nucleotide sequence that encodes an ortholog of an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said nucleotide sequence hybridizes under stringent conditions to the opposite strand of a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6;
 - (d) a nucleotide sequence that encodes a fragment of an amino acid sequence

selected from the group consisting of SEQ ID NOS:2, 5, and 7, wherein said fragment comprises at least 10 contiguous amino acids; and

- (e) a nucleotide sequence that is the complement of a nucleotide sequence of (a)-(d).
 - 6. A gene chip comprising a nucleic acid molecule of claim 5.
 - 7. A transgenic non-human animal comprising a nucleic acid molecule of claim 5.
 - 8. A nucleic acid vector comprising a nucleic acid molecule of claim 5.
 - 9. A host cell containing the vector of claim 8.
- 10. A method for producing any of the peptides of claim 1 comprising introducing a nucleotide sequence encoding any of the amino acid sequences in (a)-(d) into a host cell, and culturing the host cell under conditions in which the peptides are expressed from the nucleotide sequence.
- 11. A method for producing any of the peptides of claim 2 comprising introducing a nucleotide sequence encoding any of the amino acid sequences in (a)-(d) into a host cell, and culturing the host cell under conditions in which the peptides are expressed from the nucleotide sequence.
- 12. A method for detecting the presence of any of the peptides of claim 2 in a sample, said method comprising contacting said sample with a detection agent that specifically allows detection of the presence of the peptide in the sample and then detecting the presence of the peptide.
- 13. A method for detecting the presence of a nucleic acid molecule of claim 5 in a sample, said method comprising contacting the sample with an oligonucleotide that hybridizes to said nucleic acid molecule under stringent conditions and determining whether the oligonucleotide binds to said nucleic acid molecule in the sample.
 - 14. A method for identifying a modulator of a peptide of claim 2, said method

comprising contacting said peptide with an agent and determining if said agent has modulated the function or activity of said peptide.

- 15. The method of claim 14, wherein said agent is administered to a host cell comprising an expression vector that expresses said peptide.
- 16. A method for identifying an agent that binds to any of the peptides of claim 2, said method comprising contacting the peptide with an agent and assaying the contacted mixture to determine whether a complex is formed with the agent bound to the peptide.
- 17. A pharmaceutical composition comprising an agent identified by the method of claim 16 and a pharmaceutically acceptable carrier therefor.
- 18. A method for treating a disease or condition mediated by a human phosphatase protein, said method comprising administering to a patient a pharmaceutically effective amount of an agent identified by the method of claim 16.
- 19. A method for identifying a modulator of the expression of a peptide of claim 2, said method comprising contacting a cell expressing said peptide with an agent, and determining if said agent has modulated the expression of said peptide.

20. An isolated human phosphatase peptide having an amino acid sequence that shares at least 70% homology with an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7.

- 21. A peptide according to claim 20 that shares at least 90 percent homology with an amino acid sequence selected from the group consisting of SEQ ID NOS:2, 5, and 7.
- 22. An isolated nucleic acid molecule encoding a human phosphatase peptide, said nucleic acid molecule sharing at least 80 percent homology with a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6.
- 23. A nucleic acid molecule according to claim 22 that shares at least 90 percent homology with a nucleic acid molecule selected from the group consisting of SEQ ID NOS:1, 3, 4, and 6.

SPLICE FORM 1: 1/17

1	CGTCCTTCCT	GGTCCTGCGG	GTCCAGGACT	GTCCGCGGG	TTGAGGGAAG
51	GGGCCGTGCC	CGGTGCCAGC	CCAGGTGCTC	GCGGCCTGGC	TCCATGGCCC
101	TGGTCACAGT	GAGCCGTTCG	CCCCCGGGCA	GCGGCGCCTC	CACGCCCGTG
151	GGGCCCTGGG	ACCAGGCGGT	CCAGCGAAGG	AGTCGACTCC	AGCGAAGGCA
201	GAGCTTTGCG	GTGCTCCGTG	GGGCTGTCCT	GGGACTGCAG	GATGGAGGG
251	ACAATGATGA	TGCAGCAGAG	GCCAGTTCTG	AGCCAACAGA	GAAGGCCCCG
301	AGTGAGGAGG	AGCTCCACGG	GGACCAGACA	GACTTCGGGC	AAGGATCCCA
351	GAGTCCCCAG	AAGCAGGAGG	AGCAGAGGCA	GCACCTGCAC	CTCATGGTAC
401		GCCGCAGGAT			
451	CCCCGGCCTC	CCCGGCTCCG	CTACCTGCTG	GTAGTTTCTA	CACGAGAAGG
501		AGCCAGGATG			
551	ACAGCAGCTC	CCCCAGCTGC	ACCCTGGGCC	TGGTCTTGCC	CCTCTGGAGT
601		TGTACTTAGA			
651		CGGATCTTCA			
701		ATTGCACCAA			
751		GCAGTGCCCT			
801	GAACTCCGAA	CAGAGCTGCC	TCAATGAGTG	GACGGCTATG	GCCGACCTGG
851		GCCTCCCAGC			
901		AGGCGATCCG			
951		AGTGTCACTT			
1001		CCCCCTCCAG			
1051	CTGCTGCTGG			TCCCGCATCT	
1101	CTACCTGGGC			CCTGGAGGAG	
1151	ACAGGGTCAC	CCACATCTTG			
1201	CCTGAGCGCT			CTCTGGGATG	
1251	CCAGCTGCTG	CCGCACTGGA			
1301		CACCCACGTG			
1351		CAGTGCTGGC			
1401	GGAGCAGGCC			CCGGCCCATC	
1451	ACCCTGGCTT	CCTGCGCCAG			
1501		GGTGGTGGGG			
1551	AGGCAGCCCC	GAAAGAAGAG	CCTGGGCCAC	GGCCACGTAT	AAACCTCCGA
1601	GGGGTCATGA	GGTCCATCAG	TCTTCTGGAG	CCCTCCTTGG	AGCTGGAGAG
1651	CACCTCAGAG	ACCAGTGACA	TGCCAGAGGT	CTTCTCTTCC	CACGAGTCTT
1701	CACATGAAGA	GCCTCTGCAG	CCCTTCCCAC	AGCTTGCAAG	GACCAAGGGA
1751		TGGACAGGGG			
1801		CTCCAGGGCA			
1851	TCCAGGAGCA	GGAGCAGGGG	CAGGGGCAGG	GGCAGGGAGA	GCCCTGCATT
1901	TCCTCTACGC	CCAGGTTCCG	GAAGGTGGTG	AGACAGGCCA	GCGTGCATGA
1951	CAGTGGAGAG	GAGGGCGAGG	CCTGAGCCCT	CACACATGCC	CACGCTCCCC
2001	TGACACTGAA	GAGGATCCAC	AACTCCTTGG	AGAAACACCC	TCACGTCTGT
2051	TGCCGCACAC	ATTCCTCTCA	GCTCCGCCCC	ATACCCGTCA	CTACAGCCTC
2101	ACCTCCCACC	CCTGTCACTA	CGGCCTCACC	TCCCACCCCT	GTCACTACAG
2151	CCTCACCTCC	TACAGCCTTA	AGTCCCAGGC	CCATGTCTGC	CTGTCCAAGG
2201	GCTCAAGACT	TTCTAACTGG	GATGTGGTAG	AGGGACTGAA	GGTACCTTTG
2251	GGGGCAACAG	CACCCTAGTT	TCATTCTCAA	CTCTAGCCCT	GCACACTCAC
2301	CTGTGGCACG	GAATGAAAAC	AGAGCTTCCC	GTGCAAAAAG	GGTCACGCCT
2351	CCCACCCCCG	CCCCCTCCCT	GCACCTCCTG	TCCTCTCCCA	GTTCATTCCT
2401	GGAACCAGCC	AGGCCAGGCA	ACCAGTGGCC	CCCAAAGGCA	GGCAGGATCC
2451	TCAGGCCCCA	GCCGCGGGAG	GCTGGAAGGG	CTGGCAGATC	GCTTCCCTCA
2501	TCCACCTCCA	CCGGTCCAGG	TCTTTGCTGC	TGTCCCCAGA	CCTCCTGTGA
2551	CACCACGCCA	GATCACAGGG	CACCAGGCCA	GAGATAGTCT	TCTTTTTGTC
2601	CTTTCTGGCC	TCTGGCTAGT	CAGTTTTTCA	TAGCCTTACA	GTATCTGGCT
2651	TTGTACTGAG	AAATAAAACA	CATTTTCATA	АААААААА	AAAAAAAAA
2701	AAAA (SEQ :	ID NO:1)			

FEATURES:

5' UTR: 1-93 Start: 94-1506 Stop: 1509 3' UTR: 1510-2704

SPLICE FORM 2:

FIGURE 1A

		AAGGGGCCGT	2/	17	
1					
51	GGCTCCATGG	CCCTGGTCAC	AGTGAGCCGT	TCGCCCCCGG	GCAGCGGCGC
101	CTCCACGCCC	GTGGGGCCCT	GGGACCAGGC	GGTCCAGCGA	AGGAGTCGAC
151	TCCAGCGAAG	GCAGAGCTTT	GCGGTGCTCC	GTGGGGCTGT	CCTGGGACTG
201	CAGGATGGAG	GGGACAATGA	TGATGCAGCA	GAGGCCAGTT	CTGAGCCAAC
251	AGAGAAGGCC	CCGAGTGAGG	AGGAGCTCCA	CGGGGACCAG	ACAGACTTCG
301	GGCAAGGATC	CCAGAGTCCC	CAGAAGCAGG	AGGAGCAGAG	GCAGCACCTG
351	CACCTCATGG	TACAGCTGCT	GAGGCCGCAG	GATGACATCC	GCCTGGCAGC
401	CCAGCTGGAG	GCACCCCGGC	CTCCCCGGCT	CCGCTACCTG	CTGGTAGTTT
451	CTACACGAGA	AGGAGAAGGT	CTGAGCCAGG	ATGAGACGGT	CCTCCTGGGC
501		CTGACAGCAG			
551		AGTGACACCC			
601		TGGTGGGCAA			
651		CCACACTCCA			
701		CTTGTACCGG			=
751		ACTGAACTCC			
801		TGGAGTCTCT			
851		GAGCAGATGG			
901		CAGTGACCTG			
951		TGCGCCTGGG			
1001		ATGCTGCTGC			
1051		CCTCTACCTG			
1101		GGAACAGGGT			
1151		TACCCTGAGC			
1201		GGCCCAGCTG			
1251		CAAGAGCACA			
1301		CGCTCAGCGG			
1351		CCTGGAGCAG			
1401		CCAACCCTGG			
1451		GCCAGCCGCC			
1501		AGAGGAGCAC			
1551		CAGAACCTGA			
1601		CAGGCAGCCC			
1651		AGGGGTCATG			
1701		GCACCTCAGA			
1751		TCACATGAAG			
1801		AGGCCAGCAG			
1851		CAGTGGTTAC			
1901		TTCCAGGAGC			
1951		TTCCTCTACG			
2001		ACAGTGGAGA			
2051		CTGACACTGA			
2101		TTGCCGCACA			
2151		CACCTCCCAC			
2201		GCCTCACCTC			
2251		GGCTCAAGAC			
2301		GGGGGCAACA			
2351		CCTGTGGCAC			
2401	GGGTCACGCC	TCCCACCCC	GCCCCTCCC	TGCACCTCCT	GTCCTCTCCC
2451	AGTTCATTCC	TGGAACCAGC	CAGGCCAGGC	AACCAGTGGC	CCCCAAAGGC
2501	AGGCAGGATC	CTCAGGCCCC	AGCCGCGGGA	GGCTGGAAGG	GCTGGCAGAT
2551	CGCTTCCCTC	ATCCACCTCC	ACCGGTCCAG	GTCTTTGCTG	CTGTCCCCAG
2601		ACACCACGCC			
2651	TTCTTTTTGT	CCTTTCTGGC	CTCTGGCTAG	TCAGTTTTTC	ATAGCCTTAC
2701	AGTATCTGGC	TTTGTACTGA	GAAATAAAAC	ACATTTTCAT	AAAAAAAA
2751	AAAAAAAA	AAAAAAAAA	AAAAAAAA	AAAAAAAAA	AAAAAAAA
2801	AAAAAAAAA	AAAAAAAAA	AAAAAAAAA	AAAAAAAAA	AAAAAAAA
2851	AA (SEQ ID	NO:2)			

FEATURES:

5' UTR: 1-56 Start: 57 Stop: 2034 3' UTR: 2037-2852

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SPLICE FO	RM 3:			•	
1		CGGGTCCAGG	ACTGTCCCGC	GGGGTTGAGG	GAAGGGCCG
51		CAGCCCAGGT			
101		TTCGCCCCCG			
151		CGGTCCAGCG			
201		CGTGGGGCTG			
251		AGAGGCCAGT			
301		ACGGGGACCA			
351		GAGGAGCAGA			
401		GGATGACATC			
451		TCCGCTACCT			
501		GATGAGACGG			
551		CTGCACCCTG			
601		TAGATGGAGA			
651		TTCAAGCCCA			
701		GGAGCAGGCG			
751 751		TGGAGAGTGT			
801		GGGCTCCCCC			
851		GCTGGTGGCA			
		TGGGCTCAGA			
901					
951		GTCACCCACA			
1001		GCGCTTCACC			
1051		TGCTGCCGCA			
1101		CAGGGCACCC			
1151		GGCCACAGTG			
1201		AGGCCCTGCG			
1251		GGCTTCCTGC			
1301		CTGAGGGTGG			
1351		GCCCCGAAAG			
1401		GTCATGAGGT			
1451		CCTCAGTAGA			
1501		ACATGAAGAG			
1551		GCCAGCAGGT			
1601		GTGGTTACCC			
1651		CCAGGAGCAG			
1701		CCTCTACGCC			
1751		AGTGGAGAGG			
1801		GACACTGAAG	_		
1851		GCCGCACACA			
1901		CCTCCCACCC			
1951		CTCACCTCCT			
2001		CTCAAGACTT			
2051		GGGCAACAGC			
2101		TGTGGCACGG			
2151		CCACCCCGC			_
2201		GAACCAGCCA			
2251		CAGGCCCCAG			
2301		CCACCTCCAC			
2351		ACCACGCCAG			
2401		TTTCTGGCCT			
2451		TGTACTGAGA			
2501	AAAAAAAAA	AAAAAAAAA	AAAAAAAAA	ААААААААА	(SEQ ID NO:3)

FEATURES:

5' UTR: 1-88 Start: 88 Stop: 1311 3' UTR: 1315-2540

Homologous proteins: Top 10 BLAST Hits

4/17 Score E SPLICE FORM 1: gi|8923483|ref|NP 060327.1| hypothetical protein FLJ20515 >gi|7... 576 e-163 gi|6714641|dbj|BAA89534.1| (AB036834) MAP kinase phosphatase [D... 337 2e-91 gi|8922777|ref|NP 060746.1| hypothetical protein FLJ10928 >gi|7... 233 3e-60 qi|7242951|dbj|BAA92536.1| (AB037719) KIAA1298 protein [Homo sa... 143 4e-33 gi|7301242|gb|AAF56372.1| (AE003750) CG6238 gene product [Droso... 124 2e-27 gi|4150963|emb|CAA77232.1| (Y18620) DsPTP1 protein [Arabidopsis... 113 5e-24 gi|9294518|dbj|BAB02780.1| (AB023036) dual-specificity protein ... 113 5e-24 gi|6862915|gb|AAF30304.1|AC018907 4 (AC018907) putative dual-sp... 94 2e-18 gi|6015037|sp|054838|DUS5 RAT DUAL SPECIFICITY PROTEIN PHOSPHAT... 92 9e-18 gi|9910432|ref|NP 064570.1| mitogen-activated protein kinase ph... 90 3e-17 SPLICE FORM 2: gi|8923483|ref|NP 060327.1| hypothetical protein FLJ20515 [Homo... 576 e-163 gi|6714641|dbj|BAA89534.1| (AB036834) MAP kinase phosphatase [D... 340 6e-92 gi|8922777|ref|NP 060746.1| hypothetical protein FLJ10928 [Homo... 229 1e-58 gi|7242951|dbj|BAA92536.1| (AB037719) KIAA1298 protein [Homo sa... 162 1e-38 gi|9294518|dbj|BAB02780.1| (AB023036) dual-specificity protein ... 8e-24 113 gi|4150963|emb|CAA77232.1| (Y18620) DsPTP1 protein [Arabidopsis... 113 8e-24 gi|4758212|ref|NP 004411.1| dual specificity phosphatase 8 [Hom... 3e-19 99 gi|6679156|ref|NP 032774.1| neuronal tyrosine/threonine phospha... 96 2e-18 gi|6862915|gb|AAF30304.1|AC018907 4 (AC018907) putative dual-sp... 94 6e-18 gi|6015037|sp|054838|DUS5 RAT DUAL SPECIFICITY PROTEIN PHOSPHAT... 92 2e-17 SPLICE FORM 3: gi|8923483|ref|NP_060327.1| hypothetical protein FLJ20515 [Homo... 410 e-113 gi|8922777|ref|NP 060746.1| hypothetical protein FLJ10928 [Homo... 233 7e-60 gi|6714641|dbj|BAA89534.1| (AB036834) MAP kinase phosphatase [D... 224 5e-57 gi|7242951|dbj|BAA92536.1| (AB037719) KIAA1298 protein [Homo sa... 143 5e-33 gi|9294518|dbj|BAB02780.1| (AB023036) dual-specificity protein ... 113 1e-23 gi|4150963|emb|CAA77232.1| (Y18620) DsPTP1 protein [Arabidopsis... 113 1e-23 gi|6862915|gb|AAF30304.1|AC018907 4 (AC018907) putative dual-sp... 5e-18 94 gi|6015037|sp|054838|DUS5 RAT DUAL SPECIFICITY PROTEIN PHOSPHAT... 92 2e-17 gi|9910432|ref|NP 064570.1| mitogen-activated protein kinase ph... 90 7e-17 gi|9911130|gb|AAA64693.2| (U15932) protein phosphatase [Homo sa... 90 1e-16 BLAST to dbEST: SPLICE FORM 1:

	Score	E
gi 9807071 /dataset=dbest /taxon=960	1404	0.0
gi 10317998 /dataset=dbest /taxon=96	1316	0.0
gi 10151079 /dataset=dbest /taxon=96	1249	0.0
gi 10401153 /dataset=dbest /taxon=960	1180	0.0
gi 10329921 /dataset=dbest /taxon=96	1124	0.0
gil7632969 /dataset=dbest /taxon=960	791	0.0
gi 9155111 /dataset=dbest /taxon=9606	779	0.0
gi 10994242 /dataset=dbest /taxon=96	450	e-124

EXPRESSION INFORMATION FOR MODULATORY USE:

gi|10994242 Human ovary tumor tissue

SPLICE FORM 1:

library source:

informat	tion from BLAST dbEST hits:
	Pancreas
3 Human	colon adenocarcinoma
Human	Pancreas:adenocarcinoma
	Pancreas:epitheliod carcinoma
Human	lung: large cell carcinoma
Human	kidney: renal cell carcinoma
Human	Placenta choriocarcinoma
	Human Human Human Human Human Human

FIGURE 1D

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Expression information from PCR-based tissue screening panels:

Human Brain

Human Fetal brain

Human fetal heart

Human fetal kidney

Human heart

Human kidney

Human uterus

Human thyroid

PCT/US01/42995 WO 02/42436

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SPLICE FORM 1: 1 MALVTVSRSP PGSGASTPVG PWDQAVQRRS RLQRRQSFAV LRGAVLGLQD 51 GGDNDDAAEA SSEPTEKAPS EEELHGDQTD FGQGSQSPQK QEEQRQHLHL 101 MVQLLRPQDD IRLAAQLEAP RPPRLRYLLV VSTREGEGLS QDETVLLGVD 151 FPDSSSPSCT LGLVLPLWSD TQVYLDGDGG FSVTSGGQSR IFKPISIQTM 201 WATLOVLHOA CEAALGSGLV PGGSALTWAS HYQERLNSEQ SCLNEWTAMA 251 DLESLRPPSA EPGGSSEQEQ MEQAIRAELW KVLDVSDLES VTSKEIRQAL 301 ELRLGLPLQQ YRDFIDNQML LLVAQRDRAS RIFPHLYLGS EWNAANLEEL 351 QRNRVTHILN MAREIDNFYP ERFTYHNVRL WDEESAQLLP HWKETHRFIE 401 AARAQGTHVL VHCKMGVSRS AATVLAYAMK QYECSLEQAL RHVQELRPIA 451 RPNPGFLRQL QIYQGILTAR T (SEQ ID NO:4)

SPLICE FORM 2:

- 1 MALVTVSRSP PGSGASTPVG PWDQAVQRRS RLQRRQSFAV LRGAVLGLQD 51 GGDNDDAAEA SSEPTEKAPS EEELHGDQTD FGQGSQSPQK QEEQRQHLHL 101 MVQLLRPQDD IRLAAQLEAP RPPRLRYLLV VSTREGEGLS QDETVLLGVD 151 FPDSSSPSCT LGLVLPLWSD TQVYLDGDGG FSVTSGGQSR IFKPISIQTM 201 WATLOVLHOA CEAALGSGLV PGGSALTWAS HYQERLNSEQ SCLNEWTAMA 251 DLESLRPPSA EPGGSSEQEQ MEQAIRAELW KVLDVSDLES VTSKEIRQAL 301 ELRLGLPLQQ YRDFIDNQML LLVAQRDRAS RIFPHLYLGS EWNAANLEEL 351 ORNRVTHILN MAREIDNFYP ERFTYHNVRL WDEESAQLLP HWKETHRFIE 401 AARAQGTHVL VHCKMGVSRS AATVLAYAMK QYECSLEQAL RHVQELRPIA 451 RPNPGFLRQL QIYQGILTAS RQSHVWEQKV GGVSPEEHPA PEVSTPFPLL 501 PPEPEGGEE KVVGMEESQA APKEEPGPRP RINLRGVMRS ISLLEPSLEL 551 ESTSETSDMP EVFSSHESSH EEPLQPFPQL ARTKGGQQVD RGPQPALKSR 601 QSVVTLQGSA VVANRTQAFQ.EQEQGQGQG GEPCISSTPR FRKVVRQASV
- SPLICE FORM 3:
 - 1 MALVTVSRSP PGSGASTPVG PWDOAVORRS RLORROSFAV LRGAVLGLOD 51 GGDNDDAAEA SSEPTEKAPS EEELHGDQTD FGQGSQSPQK QEEQRQHLHL 101 MVQLLRPQDD IRLAAQLEAP RPPRLRYLLV VSTREGEGLS QDETVLLGVD
 - 151 FPDSSSPSCT LGLVLPLWSD TQVYLDGDGG FSVTSGGQSR IFKPISIQTM 201 WSSEQEQMEQ AIRAELWKVL DVSDLESVTS KEIRQALELR LGLPLQQYRD
 - 251 FIDNOMLLLV AQRDRASRIF PHLYLGSEWN AANLEELQRN RVTHILNMAR 301 EIDNFYPERF TYHNVRLWDE ESAQLLPHWK ETHRFIEAAR AQGTHVLVHC
 - 351 KMGVSRSAAT VLAYAMKQYE CSLEQALRHV QELRPIARPN PGFLRQLQIY
 - 401 QGILTART (SEQ ID NO:6)

651 HDSGEEGEA (SEQ ID NO:5)

FEATURES:

Functional domains and key regions:

SPLICE FORM 1:

[1] PDOC00004 PS00004 CAMP PHOSPHO SITE cAMP- and cGMP-dependent protein kinase phosphorylation site

34-37 RRQS

[2] PDOC00005 PS00005 PKC PHOSPHO SITE Protein kinase C phosphorylation site

Number of matches: 6

- 65-67 TEK 1
- 132-134 STR 2
- 254-256 SLR 3
- 292-294 TSK
- 395-397 THR
- 468-470 TAR

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[3] PDOC00006 PS00006 CK2 PHOSPHO_SITE Casein kinase II phosphorylation site

Number of matches: 6
1 70-73 SEEE
2 132-135 STRE
3 140-143 SQDE
4 266-269 SEQE
5 286-289 SDLE
6 292-295 TSKE

[4] PDOC00007 PS00007 TYR_PHOSPHO_SITE Tyrosine kinase phosphorylation site

363-369 REIDNFY

[5] PDOC00008 PS00008 MYRISTYL N-myristoylation site

Number of matches: 8

1 12-17 GSGAST
2 43-48 GAVLGL
3 47-52 GLQDGG
4 218-223 GLVPGG
5 223-228 GSALTW
6 339-344 GSEWNA
7 416-421 GVSRSA
8 465-470 GILTAR

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```
BLAST Alignment to Top Hit:
SPLICE FORM 1:
>gi|8923483|ref|NP_060327.1| hypothetical protein FLJ20515
          >qi|7020674|dbj|BAA91228.1| (AK000522) unnamed protein
          product [Homo sapiens]
          Length = 394
 Score = 576 \text{ bits } (1469), \text{ Expect} = e-163
 Identities = 290/312 (92%), Positives = 296/312 (93%), Gaps = 1/312 (0%)
Query: 1 MALVTVSRSPPGSGASTPVGPWDQAVQRRSRLQRRQSFAVLRGAVLGLQDGGDNDDAAEA 60
          MALVTVSRSPPGSGASTPVGPWDQAVQRRSRLQRRQSFAVLRGAVLGLQDGGDNDDAAEA
Sbjct: 1 MALVTVSRSPPGSGASTPVGPWDQAVQRRSRLQRRQSFAVLRGAVLGLQDGGDNDDAAEA 60
Query: 61 SSEPTEKAPSEEELHGDQTDFGQGSQSPQKQEEQRQHLHLMVQLLRPQDDIRLAAQLEAP 120
          SSEPTEKAPSEEELHGDQTDFGQGSQSPQKQEEQRQHLHLMVQLLRPQDDIRLAAQLEAP
Sbjct: 61 SSEPTEKAPSEEELHGDQTDFGQGSQSPQKQEEQRQHLHLMVQLLRPQDDIRLAAQLEAP 120
Query: 121 RPPRLRYLLVVSTREGEGLSQDETVLLGVDFPDSSSPSCTLGLVLPLWSDTQVYLDGDGG 180
          RPPRLRYLLVVSTREGEGLSQDETVLLGVDFPDSSSPSCTLGLVLPLWSDTQVYLDGDGG
Sbjct: 121 RPPRLRYLLVVSTREGEGLSQDETVLLGVDFPDSSSPSCTLGLVLPLWSDTQVYLDGDGG 180
Query: 181 FSVTSGGQSRIFKPISIQTMWATLQVLHQACEAALGSGLVPGGSALTWASHYQERLNSEQ 240
          FSVTSGGQSRIFKPISIQTMWATLQVLHQACEAALGSGLVPGGSALTWASHYQERLNSEQ
Sbjct: 181 FSVTSGGQSRIFKPISIQTMWATLQVLHQACEAALGSGLVPGGSALTWASHYQERLNSEQ 240
Query: 241 SCLNEWTAMADLESLRPPSAEPGGSSEQEQMEQAIRAELWKVLDV-SDLESVTSKEIRQA 299
          SCLNEWTAMADLESLRPPSAEPGGSSEQEQMEQAIRAELWKVL++ S E+
Sbjct: 241 SCLNEWTAMADLESLRPPSAEPGGSSEQEQMEQAIRAELWKVLELESTSETSDMPEVFSS 300
Query: 300 LELRLGLPLQQY 311
                PLQ +
Sbjct: 301 HESSHEEPLQPF 312 (SEQ ID NO:8)
>gi|6714641|dbj|BAA89534.1| (AB036834) MAP kinase phosphatase
           [Drosophila melanogaster]
          Length = 1045
 Score = 337 bits (854), Expect = 2e-91
 Identities = 204/537 (37%), Positives = 284/537 (51%), Gaps = 81/537 (15%)
          MALVTVSRSPPGSGA-STPVGPWDQAVQRRSRLQRRQSFAVLRGAVLGLQDGGDNDDAAE 59
Query: 1
          MALVTV RSP +G+ S G + R + F +G L L
Sbjct: 1 MALVTVQRSPSVAGSCSNSDGESEDDEGNSKGNDRSECFFAGKGTALVL------ 49
Query: 60 ASSEPTEKAPSEEELHGDQTDFGQGSQSPQKQEEQRQHLHLMVQLLRPQDDIRLAAQLEA 119
          A + SE L D T +QS + + HL M LL+ +D +++A +LE+
Sbjct: 50 ALKDIPPLTQSERRLSTDSTRSSNSTQS--NNSDIQLHLQSMFYLLQREDTLKMAVKLES 107
Sbjct: 108 QRSNRTRYLVIASRSCCRSGTSDRRRHRIMRHHSVKVGGSAGTKSSTSPAVPTQRQLSVE 167
Query: 159 -----VLPLWSDTQVY 174
                                          C LG+ V+P+ +DT ++
Sbjct: 168 QTATEASSKCDKTADKENATAAGDNKNTSGMEESCLLGIDCNERTTIGLVVPILADTTIH 227
Query: 175 LDGDGGFSVTSGGQSRIFKPISIQTMWATLQVLHQACEAALGSGLVPGGSALTWASHYQE 234
          LDGDGGFSV ++ IFKP+S+Q MW+ LQ LH+ + A + G + W S Y+
Sbjct: 228 LDGDGGFSVKVYEKTHIFKPVSVQAMWSALQTLHKVSKKARENNFYASGPSHDWLSSYER 287
```

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Query:	235	RLNSEQSCLNEWTAMADLESLRPPSAEPGGSSEQEQMEQAIRAELWKVLDVSDLESVT 292 R+ S+QSCLNEW AM LES RPPS + E+E+E I++L ++ DL+VT	
Sbjct:	288	RIESDQSCLNEWNAMDALESRRPPSPDAIRNKPPEKEETESVIKMKLKAIMMSVDLDEVT 347	
Query:	293	SKEIRQALELRLGLPLQQYRDFIDNQMLLLVAQRDRASRIFPHLYLGSEWNAANLEELQR 352 SK IR LE L + L +Y+ FID +ML+++ Q D ++IF H+YLGSEWNA+NLEELO+	
Sbjct:	348	SKYIRGRLEEILDMDLGEYKSFIDAEMLVILGQMDAPTKIFEHVYLGSEWNASNLEELQK 407	
Query:	353	NRVTHILNMAREIDNFYPERFTYHNVRLWDEESAQLLPHWKETHRFIEAARAQGTHVLVH 412 N V HILN+ REIDNF+P F Y NVR++D+E LL +W +T R+I A+A+G+ VLVH	
Sbjct:	408	NGVRHILNVTREIDNFFPGTFEYFNVRVYDDEKTNLLKYWDDTFRYITRAKAEGSKVLVH 467	
Query:	413	CKMGVSRSAATVLAYAMKQYECSLEQALRHVQELRPIARPNPGFLRQLQIYQGILTA 469 CKMGVSRSA+ V+AYAMK Y+ +QAL HV++ R +PN FL QL+ Y G+L A	
Sbjct: (SEQ]		CKMGVSRSASVVIAYAMKAYQWEFQQALEHVKKRRSCIKPNKNFLNQLETYSGMLDA 524	
>gi 892	2277	7 ref NP_060746.1 hypothetical protein FLJ10928 >gi 7023283 dbj BAA91913.1 (AK001790) unnamed protein product [Homo sapiens] Length = 141	
		233 bits (588), Expect = 3e-60 s = 111/111 (100%), Positives = 111/111 (100%)	
Query:	361	MAREIDNFYPERFTYHNVRLWDEESAQLLPHWKETHRFIEAARAQGTHVLVHCKMGVSRS 420 MAREIDNFYPERFTYHNVRLWDEESAQLLPHWKETHRFIEAARAQGTHVLVHCKMGVSRS	
Sbjct:	31	MAREIDNFYPERFTYHNVRLWDEESAQLLPHWKETHRFIEAARAQGTHVLVHCKMGVSRS 90	
Query:	421	AATVLAYAMKQYECSLEQALRHVQELRPIARPNPGFLRQLQIYQGILTART 471 AATVLAYAMKQYECSLEQALRHVQELRPIARPNPGFLRQLQIYQGILTART	
Sbict:	91	AATVLAYAMKQYECSLEQALRHVQELRPIARPNPGFLRQLQIYQGILTART 141	
(SEQ		ID	NO:10)

				10/17	
1	CGTCCTTCCT	GGTCCTGCGG	GTCCAGGACT	erceceee	TTGAGGGAAG
51	GGGCCGTGCC	CGGTGCCAGC	CCAGGTGCTC	GCGGCCTGGC	TCCATGGCCC
101	TGGTCACAGT	GAGCCGTTCG	CCCCCGGGCA	GCGGCGCCTC	CACGCCCGTG
151	GGGCCCTGGG	ACCAGGCGGT	CCAGCGAAGG	AGTCGACTCC	AGCGAAGGCA
201	GAGCTTTGCG	GTGCTCCGTG	GGGCTGTCCT	GGGACTGCAG	GATGGAGGGG
251	ACAATGATGA	TGCAGCAGAG	GCCAGTTCTG	AGCCAACAGA	GAAGGCCCCG
301	AGTGAGGAGG	AGCTCCACGG	GGACCAGACA	GACTTCGGGC	AAGGATCCCA
351	GAGTCCCCAG	AAGCAGGAGG	AGCAGAGGCA	GCACCTGCAC	CTCATGGTAC
401	AGCTGCTGAG	GCCGCAGGAT	GACATCCGCC	TGGCAGCCCA	GCTGGAGGCA
451	CCCCGGCCTC	CCCGGCTCCG	CTACCTGCTG	GTAGTTTCTA	CACGAGAAGG
501	AGAAGGTCTG	AGCCAGGATG	AGACGGTCCT	CCTGGGCGTG	GATTTCCCTG
551	ACAGCAGCTC	CCCCAGCTGC	ACCCTGGGCC	TGGTCTTGCC	CCTCTGGAGT
601	GACACCCAGG	TGTACTTAGA	TGGAGACGGG	GGCTTCAGCG	TGACGTCTGG
651	TGGGCAAAGC	CGGATCTTCA	AGCCCATCTC	CATCCAGACC	ATGTGGGCCA
701	CACTCCAGGT	ATTGCACCAA	GCATGTGAGG	CAGCTCTAGG	CAGCGGCCTT
751	GTACCGGGTG	GCAGTGCCCT	CACCTGGGCC	AGCCACTACC	AGGAGAGACT
801	GAACTCCGAA	CAGAGCTGCC	TCAATGAGTG	GACGGCTATG	GCCGACCTGG
851	AGTCTCTGCG	GCCTCCCAGC	GCCGAGCCTG	GCGGGTCCTC	AGAACAGGAG
901	CAGATGGAGC	AGGCGATCCG	TGCTGAGCTG	TGGAAAGTGT	TGGATGTCAG
951	TGACCTGGAG	AGTGTCACTT	CCAAAGAGAT	CCGCCAGGCT	CTGGAGCTGC
1001	GCCTGGGGCT	CCCCCTCCAG	CAGTACCGTG	ACTTCATCGA	CAACCAGATG
1051	CTGCTGCTGG	TGGCACAGCG	GGACCGAGCC	TCCCGCATCT	TCCCCCACCT
1101	CTACCTGGGC		ACGCAGCAAA	CCTGGAGGAG	CTGCAGAGGA
1151	ACAGGGTCAC	CCACATCTTG	AACATGGCCC	GGGAGATTGA	CAACTTCTAC
1201	CCTGAGCGCT	TCACCTACCA	CAATGTGCGC	CTCTGGGATG	AGGAGTCGGC
1251	CCAGCTGCTG	CCGCACTGGA	AGGAGACGCA	CCGCTTCATT	GAGGCTGCAA
1301	GAGCACAGGG	CACCCACGTG	CTGGTCCACT	GCAAGATGGG	CGTCAGCCGC
1351	TCAGCGGCCA	CAGTGCTGGC	CTATGCCATG	AAGCAGTACG	AATGCAGCCT
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	GGCCAGCAGG				
	AGTGGTTACC				
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1951	CAGTGGAGAG	GAGGGCGAGG	CCTGAGCCCT	CACACATGCC	CACGCTCCCC
2001	TGACACTGAA	GAGGATCCAC	AACTCCTTGG	AGAAACACCC	TCACGTCTGT
	TGCCGCACAC				
	ACCTCCCACC				
	CCTCACCTCC				
	GCTCAAGACT				
	GGGGCAACAG				
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	GGAACCAGCC				
	TCAGGCCCCA				
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	CACCACGCCA				
	CTTTCTGGCC				
	TTGTACTGAG				
	AAAA (SEQ 1				
	- ,	· - · •			

FEATURES:

Start: 94 Exon: 94-1506 Stop: 1507

CHROMOSOME MAP POSITION:

Bac accession number: AP001885

Chromosome #: 11

11/17

DNA				Protein		
Position	Major	Minor	Domain	Position	Major	Minor
577	G	A	Exon	162	G	S
1451	G	Α	Exon	453	S	N
2641	G	Α	Beyond ORF(3')			

Context:

DNA

Position

577

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GCAAGATGGGCGTCAGCCGCTCAGCGGCCACAGTGCTGCCCTATGCCATGAAGCAGTACG
AATGCAGCCTGGAGCAGGCCCTGCGCCACGTGCAGGAGCTCCGCCCCCCA
[G, A]

CCCTGGCTTCCTGCGCCAGCTGCAGATCTACCAGGGCATCCTGACGGCCAGAACCTGAGG GTGGTGGGGAGAGAGGTTGTAGGCATGGAAGAGAGCCCAGGCAGCCCCGAAAGAAGAGC CTGGGCCACGGCCACGTATAAACCTCCGAGGGGTCATGAGGTCCATCAGTCTTCTGGAGC CCTCCTTGGAGCTGGAGAGCACCTCAGAGACCAGTGACATGCCAGAGGTCTTCTCTCCC ACGAGTCTTCACATGAAGAGCCTCTGCAGCCCTTCCCACAGCTTGCAAGGACCAAGGGAG

GENE STRUCTURE MODEL:

SPLICE FORM 2	5'-e1-e2-e3-e4-e5-e6-e7-e8-e9-e10-e11-e12-e13-e14-3'
SPLICE FORM 1	5'-e1-e2-e3-e4-e5-e6-e7-e8-e9-e10-e11-e123'
SPLICE FORM 3	5'-e1-e2-e3-e4-e5-e6e8-e9-e10-e11-e123'

MULTIPLE ALIGNMENT OF CONA SEQUENCES:

SPLICE	FORM	2	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~TG	GTTGAGGGAA	
SPLICE	FORM	1	CGTCCTTCCT	GGTCCTGCGG	GTCCAGGACT	GT.CCGCGGG	GTTGAGGGAA	
SPLICE	FORM	3	~~~~~CCT	GGTCCTGCGG	GTCCAGGACT	GTCCCGCGGG	GTTGAGGGAA	
			51				100	
		_		_				

SPLICE	FORM	2	GGGGCCGTGC	CCGGTGCCAG	CCCAGGTGCT	CGCGGCCTGG	CTCCATGGCC
SPLICE	FORM	1	GGGGCCGTGC	CCGGTGCCAG	CCCAGGTGCT	CGCGGCCTGG	CTCCATGGCC
SPLICE	FORM	3	GGGGCCGTGC	CCGGTGCCAG	CCCAGGTGCT	CGCGGCCTGG	CTCCATGGCC

101 150

SPLICE FORM 2 CTGGTCACAG TGAGCCGTTC GCCCCCGGGC AGCGGCGCCT CCACGCCCGT SPLICE FORM 1 CTGGTCACAG TGAGCCGTTC GCCCCCGGGC AGCGGCGCCT CCACGCCCGT

SPLICE FORM 3	CTGGTCACAG TGAGCCGTT	c sccccessc aggscscct ccacscccst
	151	200
	GGGGCCCTGG GACCAGGCG	G TCCAGCGAAG GAGTCGACTC CAGCGAAGGC
		G TCCAGCGAAG GAGTCGACTC CAGCGAAGGC G TCCAGCGAAG GAGTCGACTC CAGCGAAGGC
	201	250
		T GGGGCTGTCC TGGGACTGCA GGATGGAGGG T GGGGCTGTCC TGGGACTGCA GGATGGAGGG
		T GGGGCTGTCC TGGGACTGCA GGATGGAGGG
	251	. 300
		A GGCCAGTTCT GAGCCAACAG AGAAGGCCCC A GGCCAGTTCT GAGCCAACAG AGAAGGCCCC
SPLICE FORM 3	GACAATGATG ATGCAGCAG	A GGCCAGTTCT GAGCCAACAG AGAAGGCCCC
CDITCE FORM A	301	350
		G GGGACCAGAC AGACTTCGGG CAAGGATCCC G GGGACCAGAC AGACTTCGGG CAAGGATCCC
SPLICE FORM 3	GAGTGAGGAG GAGCTCCAC	G GGGACCAGAC AGACTTCGGG CAAGGATCCC
CDITCE ECDM 2	351	400
		G GAGCAGAGGC AGCACCTGCA CCTCATGGTA G GAGCAGAGGC AGCACCTGCA CCTCATGGTA
SPLICE FORM 3	AGAGTCCCCA GAAGCAGGA	G GAGCAGAGGC AGCACCTGCA CCTCATGGTA
CDITCE FORM 2	401	450
		A TGACATCCGC CTGGCAGCCC AGCTGGAGGC A TGACATCCGC CTGGCAGCCC AGCTGGAGGC
SPLICE FORM 3	CAGCTGCTGA GGCCGCAGG	A TGACATCCGC CTGGCAGCCC AGCTGGAGGC
CDITCE CODM 2	451	500 C GCTACCTGCT GGTAGTTTCT ACACGAGAAG
		C GCTACCTGCT GGTAGTTTCT ACACGAGAAG C GCTACCTGCT GGTAGTTTCT ACACGAGAAG
SPLICE FORM 3	ACCCCGGCCT CCCCGGCTC	C GCTACCTGCT GGTAGTTTCT ACACGAGAAG
CDITCE FORM 2	501	550
		T GAGACGGTCC TCCTGGGCGT GGATTTCCCT T GAGACGGTCC TCCTGGGCGT GGATTTCCCT
SPLICE FORM 3	GAGAAGGTCT GAGCCAGGA	T GAGACGGTCC TCCTGGGCGT GGATTTCCCT
SPLICE FORM 2	551	600 G CACCCTGGGC CTGGTCTTGC CCCTCTGGAG
		G CACCCIGGGC CIGGICITGC CCCICIGGAG G CACCCIGGGC CIGGICITGC CCCICIGGAG
SPLICE FORM 3	GACAGCAGCT CCCCCAGCT	G CACCCTGGGC CTGGTCTTGC CCCTCTGGAG
SPLICE FORM 2	601	650 G ATGGAGACGG GGGCTTCAGC GTGACGTCTG
		G ATGGAGACGG GGGCTTCAGC GTGACGTCTG
SPLICE FORM 3	TGACACCCAG GTGTACTTA	G ATGGAGACGG GGGCTTCAGC GTGACGTCTG
SPLICE FORM 2	651	700 C AAGCCCATCT CCATCCAGAC CATGTGGGCC
		C AAGCCCATCT CCATCCAGAC CATGTGGGCC
SPLICE FORM 3	GTGGGCAAAG CCGGATCTT	C AAGCCCATCT CCATCCAGAC CATGT
SPIJICE FORM 2	701	750 A AGCATGTGAG GCAGCTCTAG GCAGCGGCCT
SPLICE FORM 1	ACACTCCAGG TATTGCACC	A AGCATGTGAG GCAGCTCTAG GCAGCGGCCT
SPLICE FORM 3		• • • • • • • • • • • • • • • • • • • •
SPLICE FORM ?	751	800 C TCACCTGGGC CAGCCACTAC CAGGAGAGAC
SPLICE FORM 1	TGTACCGGGT GGCAGTGCC	C TCACCTGGGC CAGCCACTAC CAGGAGAGAC
SPLICE FORM 3	•••••••	

13/17

			15/17					
			801				850	
SPLICE	FORM	2	TGAACTCCGA	ACAGAGCTGC	CTCAATGAGT	GGACGGCTAT	GGCCGACCTG	
SPLICE	FORM	1	TGAACTCCGA	ACAGAGCTGC	CTCAATGAGT	GGACGGCTAT	GGCCGACCTG	
SPLICE	FORM	3						
			851				900	
SPLICE	FORM	2	GAGTCTCTGC	GGCCTCCCAG	CGCCGAGCCT	GGCGGGTCCT		
			GAGTCTCTGC					
SPLICE		_	GAGICICIGC					
SPLICE	FORM	3	• • • • • • • • •	• • • • • • • • •	• • • • • • • • •	GGTCCT	CAGAACAGGA	
							950	
ODT TOP	70DM	^	901	ON COCON MOC	CMCCMCA CCM	COCCAAACOC		
			GCAGATGGAG			•		
			GCAGATGGAG					
SPLICE	FORM	3	GCAGATGGAG	CAGGUGATUU	GTGCTGAGCT	GIGGAAAGIG	TTGGATGTCA	
							1000	
			951				1000	
			GTGACCTGGA					
			GTGACCTGGA					
SPLICE	FORM	3	GTGACCTGGA	GAGTGTCACT	TCCAAAGAGA	TCCGCCAGGC	TCTGGAGCTG	
		,	1001				1050	
SPLICE	FORM	2	CGCCTGGGGC	TCCCCCTCCA	GCAGTACCGT	GACTTCATCG	ACAACCAGAT	
SPLICE	FORM	1	CGCCTGGGGC	TCCCCCTCCA	GCAGTACCGT	GACTTCATCG	ACAACCAGAT	
SPLICE	FORM	3	CGCCTGGGGC	TCCCCCTCCA	GCAGTACCGT	GACTTCATCG	ACAACCAGAT	
			1051				1100	
SPLICE	FORM	2	GCTGCTGCTG	GTGGCACAGC	GGGACCGAGC	CTCCCGCATC	TTCCCCCACC	
SPLICE	FORM	1	GCTGCTGCTG	GTGGCACAGC	GGGACCGAGC	CTCCCGCATC	TTCCCCCACC	
SPLICE	FORM	3	GCTGCTGCTG	GTGGCACAGC	GGGACCGAGC	CTCCCGCATC	TTCCCCCACC	
		•						
			1101				1150	
SPLICE	FORM	2	TCTACCTGGG	CTCAGAGTGG	AACGCAGCAA	ACCTGGAGGA	GCTGCAGAGG	
			TCTACCTGGG					
			TCTACCTGGG					
DIDIOD	10141	•	1014001000	010/10/10100	1210001100121	1100100110011	001001101100	
			1151				1200	
SDLTCE	FORM	2	AACAGGGTCA	C CC <u>D</u> C <u>D</u> TCTT	CAACATCCCC	ССССАСАТТС		
			AACAGGGTCA					
			AACAGGGTCA		=			
SEDICE	rorda	,	MCMOGGICA	CCCACATCIT	GAACATGGCC	CGGGHGAIIG	ACAMOTICIA	
			1201				1250	
CDITOR	₽ОВМ	2	CCCTGAGCGC	中中でみでで申れてて	አርአአጥርጥርርር	- CC中で中ででで表面		
			CCCTGAGCGC					
-	-							
SPLICE	FORM	3	CCCTGAGCGC	TTCACCTACC	ACAATGTGCG	CCTCTGGGAT	GAGGAGTCGG	
			1051				1200	
an tan	2001	_	1251	CCCCCT CTCC	3 3 CC3 C3 CCC	3 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	1300	
			CCCAGCTGCT					
			CCCAGCTGCT					
SPLICE	FORM	3	CCCAGCTGCT	GCCGCACTGG	AAGGAGACGC	ACCGCTTCAT	TGAGGCTGCA	
			1001				1250	
		_ ,	1301				1350	
			AGAGCACAGG					
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SPLICE	FORM	3	AGAGCACAGG	GCACCCACGT	GCTGGTCCAC	TGCAAGATGG	GCGTCAGCCG	
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SPLICE	FORM	2	CTCAGCGGCC	ACAGTGCTGG	CCTATGCCAT	GAAGCAGTAC	GAATGCAGCC	
SPLICE	FORM	1	CTCAGCGGCC	ACAGTGCTGG	CCTATGCCAT	GAAGCAGTAC	GAATGCAGCC	
SPLICE	FORM	3	CTCAGCGGCC	ACAGTGCTGG	CCTATGCCAT	GAAGCAGTAC	GAATGCAGCC	
			1401		•		1450	
SPLICE	FORM	2	TGGAGCAGGC	CCTGCGCCAC	GTGCAGGAGC	TCCGGCCCAT	CGCCCGCCCC	
			TGGAGCAGGC					
			TGGAGCAGGC					
				222333310				

14/17 SPLICE FORM 2 AACCCTGGCT TCCTGCGCCA GCTGCAGATC TACCAGGGCA TCCTGACGGC SPLICE FORM 1 AACCCTGGCT TCCTGCGCCA GCTGCAGATC TACCAGGGCA TCCTGACG.. SPLICE FORM 3 AACCCTGGCT TCCTGCGCCA GCTGCAGATC TACCAGGGCA TCCTGACG.. 1501 1550 SPLICE FORM 2 CAGCCGCCAG AGCCATGTCT GGGAGCAGAA AGTGGGTGGG GTCTCCCCAG 1551 SPLICE FORM 2 AGGAGCACCC AGCCCCTGAA GTCTCTACAC CATTCCCACT TCTTCCGCCA SPLICE FORM 1GCCA SPLICE FORM 3GCCA 1601 SPLICE FORM 2' GAACCTGAGG GTGGTGGGGA GGAGAAGGTT GTAGGCATGG AAGAGAGCCA SPLICE FORM 1 GAACCTGAGG GTGGTGGGGA GGAGAAGGTT GTAGGCATGG AAGAGAGCCA SPLICE FORM 3 GAACCTGAGG GTGGTGGGGA GGAGAAGGTT GTAGGCATGG AAGAGAGCCA 1651 SPLICE FORM 2 GGCAGCCCG AAAGAAGAGC CTGGG..CCA CGGCCACGTA TAAACCTCCG SPLICE FORM 1 GGCAGCCCCG AAAGAAGAGC CTGGG..CCA CGGCCACGTA TAAACCTCCG SPLICE FORM 3 GGCAGCCCG AAAGAAGAGC CTGGGGCCAC GGGGCACGTA TAAACCTCCG SPLICE FORM 2 AGGGGTCATG AGGTCCATCA GTCTTCTGGA GCCCTCCTT. GGAGCTGGAG SPLICE FORM 1 AGGGGTCATG AGGTCCATCA GTCTTCTGGA GCCCTCCTT. GGAGCTGGAG SPLICE FORM 3 AGGGGTCATG AGGTCCATCA GTCTTCTGGA GCCCTCCTTG GGAGCTGGAG 1751 1800 SPLICE FORM 2 AGCACCTCAG .AGACCAGTG ACATGCCAGA GGTCTTCTCT TCCCACGAGT SPLICE FORM 1 AGCACCTCAG .AGACCAGTG ACATGCCAGA GGTCTTCTCT TCCCACGAGT SPLICE FORM 3 AGCACCTCAG TAGACCAGTG ACATGCCAGA GGTCTTCTCT TCCCACGAGT 1801 SPLICE FORM 2 CTTCACATGA AGAGCCTCTG CAGCCCTTCC CACAGCTTGC AAGGACCAAG SPLICE FORM 1 CTTCACATGA AGAGCCTCTG CAGCCCTTCC CACAGCTTGC AAGGACCAAG SPLICE FORM 3 CTTCACATGA AGAGCCTCTG CAGCCCTTCC CACAGCTTGC AAGGACCAAG SPLICE FORM 2 GGAGGCCAGC AGGTGGACAG GGGGCCTCAG CCTGCCCTGA AGTCCCGCCA SPLICE FORM 1 GGAGGCCAGC AGGTGGACAG GGGGCCTCAG CCTGCCCTGA AGTCCCGCCA SPLICE FORM 3 GGAGGCCAGC AGGTGGACAG GGGGCCTCAG CCTGCCCTGA AGTCCCGCCA 1901 SPLICE FORM 2 GTCAGTGGTT ACCCTCCAGG GCAGTGCCGT GGTGGCCAAC CGGACCCAGG SPLICE FORM 1 GTCAGTGGTT ACCCTCCAGG GCAGTGCCGT GGTGGCCAAC CGGACCCAGG SPLICE FORM 3 GTCAGTGGTT ACCCTCCAGG GCAGTGCCGT GGTGGCCAAC CGGACCCAGG SPLICE FORM 2 CCTTCCAGGA GCAGGAGCAG GGGCAGGGC AGGGGCAGGG AGAGCCCTGC SPLICE FORM 1 CCTTCCAGGA GCAGGAGCAG GGGCAGGGGC AGGGCCAGGG AGAGCCCTGC SPLICE FORM 3 CCTTCCAGGA GCAGGAGCAG GGGCAGGGC AGGGGCAGGG AGAGCCCTGC 2001 2050 SPLICE FORM 2 ATTTCCTCTA CGCCCAGGTT CCGGAAGGTG GTGAGACAGG CCAGCGTGCA SPLICE FORM 1 ATTTCCTCTA CGCCCAGGTT CCGGAAGGTG GTGAGACAGG CCAGCGTGCA SPLICE FORM 3 ATTTCCTCTA CGCCCAGGTT CCGGAAGGTG GTGAGACAGG CCAGCGTGCA 2051 SPLICE FORM 2 TGACAGTGGA GAGGAGGGCG AGGCCTGAGC CCTCACACAT GCCCACGCTC SPLICE FORM 1 TGACAGTGGA GAGGAGGGCG AGGCCTGAGC CCTCACACAT GCCCACGCTC SPLICE FORM 3 ΨGACACTCCA GACCACCCCC ბርСССФСЪСС ССФСАСАСТА ССССАСССФС 2101 2150

					15/17		
					CACAACTCCT	TGGAGAAACA	 -
					CACAACTCCT CACAACTCCT		
			2151				2200
			TGTTGCCGCA		TCAGCTCCGC		TCACTACAGC
SPLICE		_			TCAGCTCCGC TCAGCTCCGC	=	- -
	10141	J		0.00.00.00	10/10010000	00011110000	
SPLICE	FORM	2	2201 CTCACCTCCC	ACCCCTGTCA	CTACGGCCTC	ACCTCCCACC	2250 CCTGTCACTA
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SPLICE	FORM	3	CTCACCTCCC	ACCCCTGTCA	CTACGGCCTC	ACCTCCCACC	CCTGTCACTA
		_	2251				2300
					TTAAGTCCCA TTAAGTCCCA		
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			2301				2350
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					TGGGATGTGG		
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451 = 4m		_	2351				2400
					GTTTCATTCT GTTTCATTCT		
					GTTTCATTCT		
			2401				2450
SPLICE	FORM	2	- _	ACGGAATGAA	AACAGAGCTT	CCCGTGCAAA	AAGGGTCACG
					AACAGAGCTT		
SPLICE	FORM	3	CACCTGTGGC	ACGGAATGAA	AACAGAGCTT	CCCGTGCAAA	AAGGGTCACG
		•	2451				2500
					CCTGCACCTC		
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			2501				2550
					GCAACCAGTG		GCAGGCAGGA
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SPLICE	FORM	3	CCIGGAACCA	GCCAGGCCAG	GCANCCAGTG	GCCCCCAAAG	GCAGGCAGGA
CDITOR	FORM	2	2551	00200000	CD CCCMCCD A	CCCCMCCCA C	2600
					GAGGCTGGAA GAGGCTGGAA		
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			2601				2650
					AGGTCTTTGC		
					AGGTCTTTGC AGGTCTTTGC		
SEDICE	FORM	3	TCATCCACCT	CCACCGGICC	AGGICITIGC	IGCIGICCCC	AGACCICCIG
QDT TOP	FODM	2	2651	CCDCDDCDCDCD	GGGCACCAGG	ここれ これ これのみ こ	2700
					GGGCACCAGG		
_					GGGCACCAGG		
			2701				2750
					AGTCAGTTTT		ACAGTATCTG
					AGTCAGTTTT AGTCAGTTTT		ACAGTATCTG ACAGTATCTG
		_		301010001			
SPLTCE	р∩рм	2	2751 CCTTTCTACT	CDCDDDTTDD	ACACATTTTC	<u> </u> עאממממשת	2800 ממממממממ
OFTICE	FORM	4	GOTTIGIACI	OUGUMU I WWW	HUNGHIIIU	WHUNDUNHAH	UNUVULARIAN!

						16117			
	SPLICE	FORM	1	GCTTTGTACT	GAGAAATAAA	ACACATTTC	АТААААААА	ААААААААА	
	SPLICE	FORM	3	GCTTTGTACT	GAGAAATAAA	ACACATTTTC	ATATTTGGTT	AAAAAAAA	
				2801				2850	
			_		AAAAAAAAA				
					~~~~~~~~				
	SPLICE	FORM	3	AAAAAAAAA	AAAAAAAA	AAAAAAAAA	AAAA~~~~	~~~~~~~	
				2051				2894	
	CDITCE	Magar	2	2851	ААААААААА	****	אראה אראה א א א		
					~~~~~~				
				~~~~~~~		~~~~~~~	~~~~~~~	~~~~	
	OLDICE	LOIGI	,						
•									
	MULTIPI	E AL	[G1	MENT OF PE	TIDE SEQUE	NCES:			
	SPLICE	FORM	2	MALVTVSRSP	PGSGASTPVG	PWDQAVQRRS	${\tt RLQRRQSFAV}$	LRGAVLGLQD	
					PGSGASTPVG				
	SPLICE	FORM	3	MALVTVSRSP	PGSGASTPVG	PWDQAVQRRS	RLQRRQSFAV	LRGAVLGLQD	
			_	51	000000000000000000000000000000000000000		Economor.	100	
					SSEPTEKAPS				
					SSEPTEKAPS				
	SPLICE	FORM	3	GGUNDUAALA	SSEPTEKAPS	FFFTUGDOLD	rGQGSQSPQK	<b>Ö</b> EEÖKÖUPUT	
				101				150	
	SPLICE	FORM	2		IRLAAQLEAP	RPPRI.RYI.I.V	VSTREGEGIS		
					IRLAAQLEAP			- <del>-</del>	
					IRLAAQLEAP				
				~ ~	~ ~				
				151				200	
					LGLVLPLWSD				
					LGLVLPLWSD	-	_	_	
	SPLICE	FORM	3	FPDSSSPSCT	LGLVLPLWSD	TQVYLDGDGG	FSVTSGGQSR	IFKPISIQTM	
			_	201				250	
					CEAALGSGLV				
					CEAALGSGLV		•		
	SPLICE	FORM	J	w	• • • • • • • • • • • • • • • • • • • •	• • • • • • • • • •	• • • • • • • • • •	• • • • • • • • •	
				251				300	
	SPLICE	FORM	2		EPGGSSEQEQ	MEGATRAELW	KVLDVSDLES		
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					SSEQEQ				
				301				350	
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					YRDFIDNOML				
	SPLICE	FORM	3	ELRLGLPLQQ	YRDFIDNQML	LLVAQRDRAS	RIFPHLYLGS	EWNAANLEEL	
				261			•	400	
	CDI TOT	FODY	^	351	MADETONE	ED EMVISION	MDDDDOSOTTO	400	
					MAREIDNFYP MAREIDNFYP				
					MAREIDNEYP				
	STHICE	PORM	J	ÖVMVATUTDM	PHAREIDNETE	EKTIIMVKD	MORESAGE	HWKETHKE IE	
				401				450	
	SPLICE	FORM	2		VHCKMGVSRS	AATVLAYAMK	OYECSLEOAL		
					VHCKMGVSRS				
					VHCKMGVSRS				
				451				500	
					QIYQGILTAS				
	SPLICE	FORM	1	RPNPGFLRQL	QIYQGILTAR	T~~~~~~	~~~~~~~	~~~~~~~	
	SPLICE	FORM	3	RPNPGFLRQL	QIYQGILTAR	T~~~~~~	~~~~~~~	~~~~~~~	
				F.0.1					
	CDITOR	EOP16	2	501	MINICHIPP AAS	A DICTION OF THE	DINITO	550	
	SEPTCE	r okm	2	rrereddet	KVVGMEESQA	AFREEFGFKP	KINTKGAWKS	TOTTELOTET	

SPLICE	FORM	1	~~~~~	~~~~~~~~~	17/17	~~~~~~	~~~~~~~~
SPLICE			~~~~~~~		~~~~~~	~~~~~~	~~~~~~~
			551				600
SPLICE	FORM	2	ESTSETSDMP	EVFSSHESSH	EEPLQPFPQL	ARTKGGQQVD	RGPQPALKSR
SPLICE	FORM	1	~~~~~~	~~~~~~~	~~~~~~	~~~~~~~	~~~~~~~~
SPLICE	FORM	3	~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~
			601				650
SPLICE	FORM	2	QSVVTLQGSA	<b>VVANRTQAFQ</b>	EQEQGQGQGQ	GEPCISSTPR	FRKVVRQASV
SPLICE	FORM	1	~~~~~~	~~~~~~~	~~~~~~	~~~~~~	~~~~~~~
SPLICE	FORM	3	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~	~~~~~~~
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SPLICE	FORM	3	~~~~~~				

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Phe Gly Gln Gly Ser Gln Ser Pro Gln Lys Gln Glu Glu Gln Arg Gln
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His Leu His Leu Met Val Gln Leu Leu Arg Pro Gln Asp Asp Ile Arg
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Leu Ala Ala Gln Leu Glu Ala Pro Arg Pro Pro Arg Leu Arg Tyr Leu
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Leu Val Val Ser Thr Arg Glu Gly Glu Gly Leu Ser Gln Asp Glu Thr
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